Physiopathologie des infections osseuses chroniques (biofilm, persistance intra-cellulaire)

JP Bru

Maladies Infectieuses

Centre Hospitalier Annecy Genevois



Inoculation
Infection aiguë
Destruction osseuse
Persistance bactérienne

Bactérie

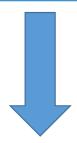
Os / matériel

Hôte Réponse immune



Inoculation
Infection aiguë
Destruction osseuse
Persistance bactérienne

Comportement bactérien

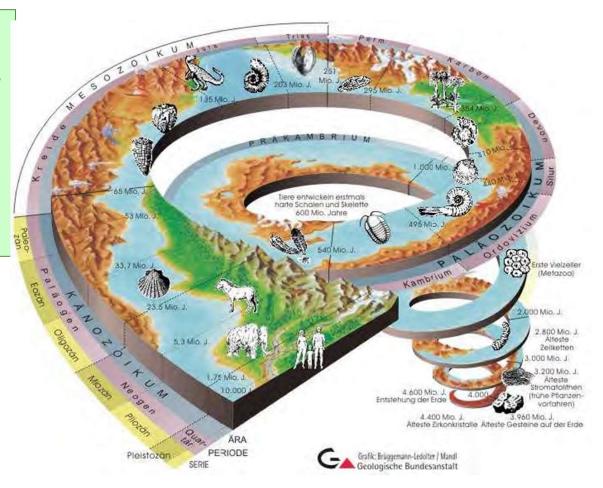


Conséquences pour infections osseuses / sur matériel (évolutives et thérapeutiques)

1/ Adhérence et organisations bactériennes

2/ Variants microcolonies

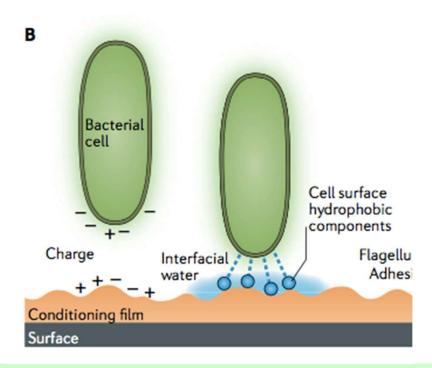
3/ Persistance intra cellulaire



Microcolonies formant biofilm identifiées :

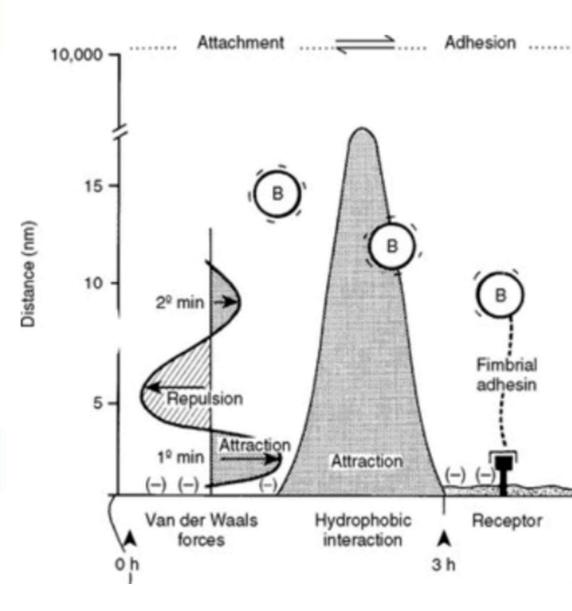
- 3.3–3.4 milliard d'années South African Kornberg formation
- 3.2-milliard d'années deep-sea hydrothermal rocks of the Pilbara Craton, Australia

Adhérence



Cécile Berne NATURE Reviews | Microobiology volume 16 | OCTOBER 2018 | 617

Gristina AG: Science 237:1588-1595, 1987



Adhérence

Flagellum Repression of flagellar rotation Adhesin production Flagellar motor Adhesin production Rotor-dependent surface sensing -Pilus Adhesin production Pilus retraction Cécile Berne

Surface

Adhesion forces

Deformation of

the cell envelope

Adhesin production

Adhesin-

NATURE Reviews | Microobiology volume 16 | OCTOBER 2018 | **617**

Adhérence & organisation bactérienne

INFECTION AND IMMUNITY, July 1982, p. 318-326 0019-9567/82/070318-09\$02.00/0

Vol. 37, No. 1

Adherence of Slime-Producing Strains of Staphylococcus epidermidis to Smooth Surfaces

GORDON D. CHRISTENSEN, 1* W. ANDREW SIMPSON, $^{1.2}$ ALAN L. BISNO, 1 AND EDWIN H. BEACHEY $^{1.2}$

320 CHRISTENSEN ET AL.

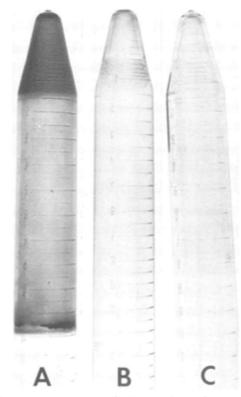
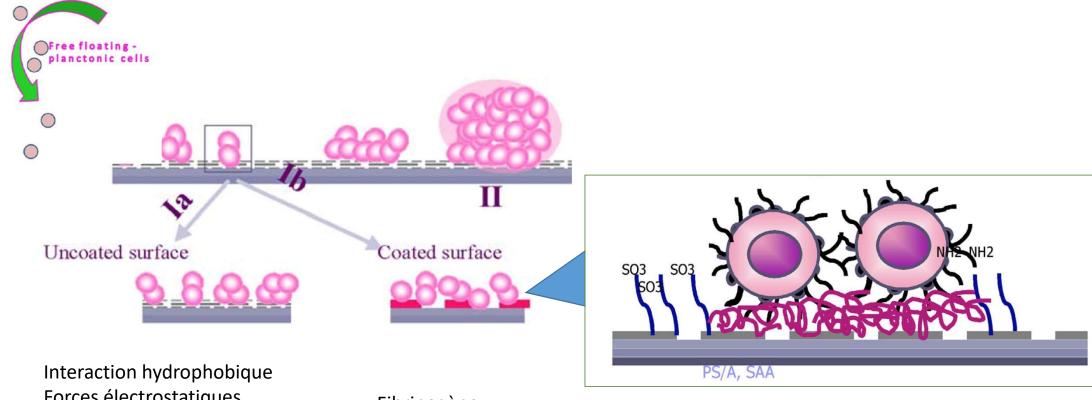


FIG. 1. Adherence of S. epidermidis grown in broth to the walls of plastic test tubes. The test tubes were emptied and stained with trypan blue. (A) Strong slime-producing strain in TSB; (B) weak slime-producing strain in TSB; (C) strong slime-producing strain grown in saccharide-free basal medium.

Adhérence & organisation bactérienne

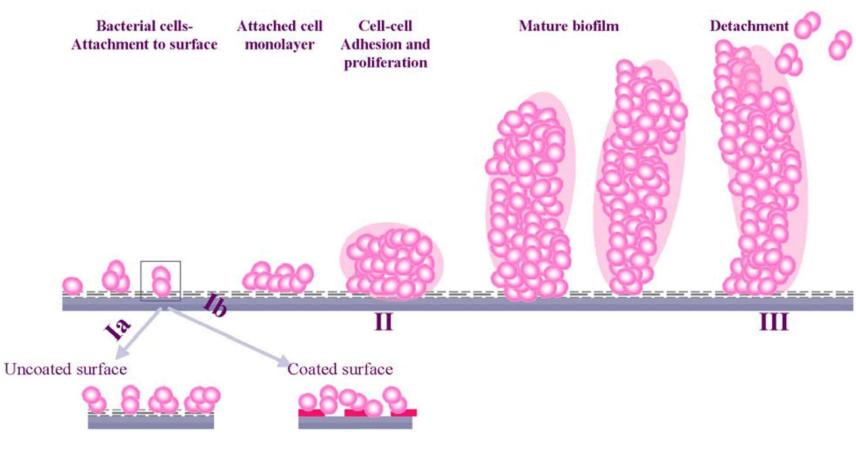


Forces électrostatiques Forces de Van der Wals

Fibrinogène facteur de Willbrand Plaquettes prothrombine

European cells & Materials Vol 8 2004 p 37-57

Formation du biofilm



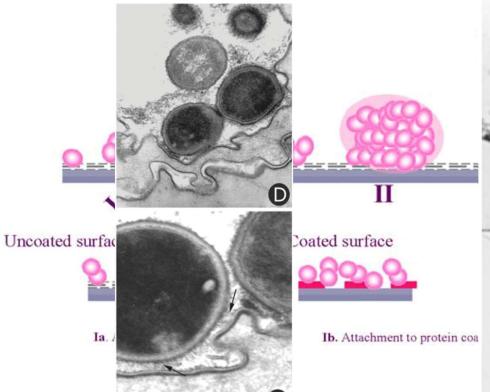
Ia. Attachment to uncoated surface by physicochemical forces Ib. Attachment to protein coated surface

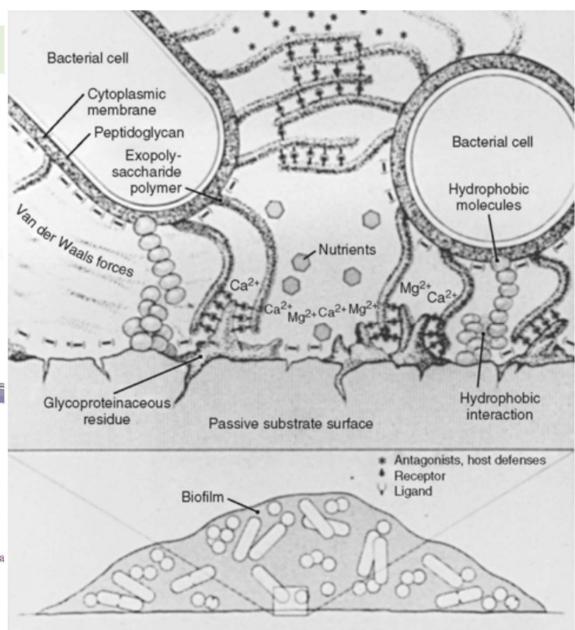
II. Cell-cell proliferation To mature biofilm III. Detachment

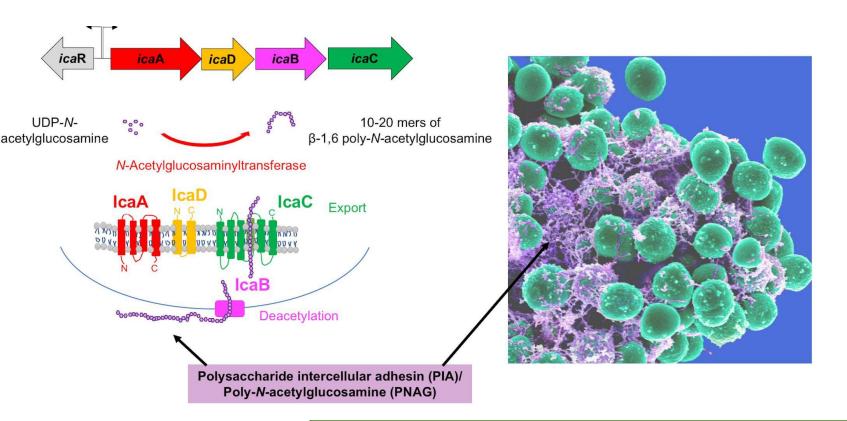
European cells & Materials Vol 8 2004 p 37-57

Formation du biofilm

Bacterial cells-Attachment to surface Attached cell monolayer Cell-cell Adhesion and proliferation







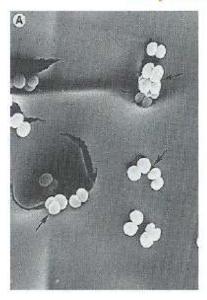
Matrice du biofilm

polysaccharide intercellular adhesion (PIA) exopolysaccharide, proteins such as accumulation-associated protein (Aap) extracellular matrix binding protein (Embp), teichoic acids, extracellular DNA (eDNA).

Channels in the biofilm are formed by Phenol-soluble modulins (PSMs)

Formation d'un biofilm

Etapes de la colonisation du matériel par S. epidermidis







2 h

Fixation des staph sur des irrégularités à la surface du matériel

4 h

Début de fabrication du "slime"

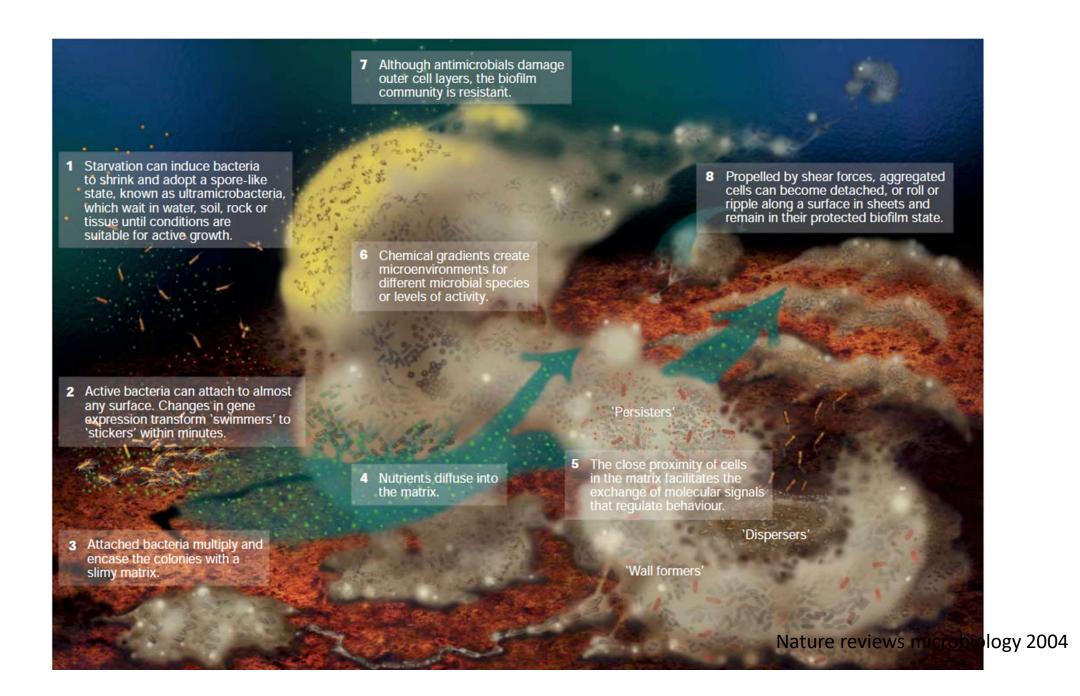
8 h

La surface du matériel est recouverte par une couche épaisse de "slime"

24 h

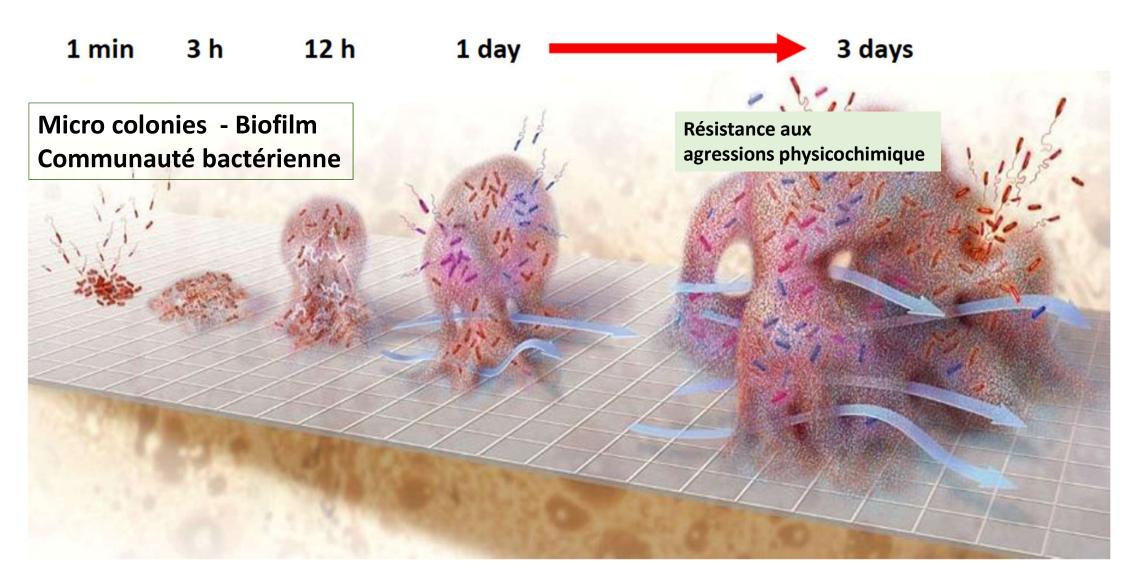
Des bactéries émergent du biofilm, libres et prêtes à se fixer ailleurs

Olson ME et al. J Biomed Mater Res, 1988



Biofilm rôle

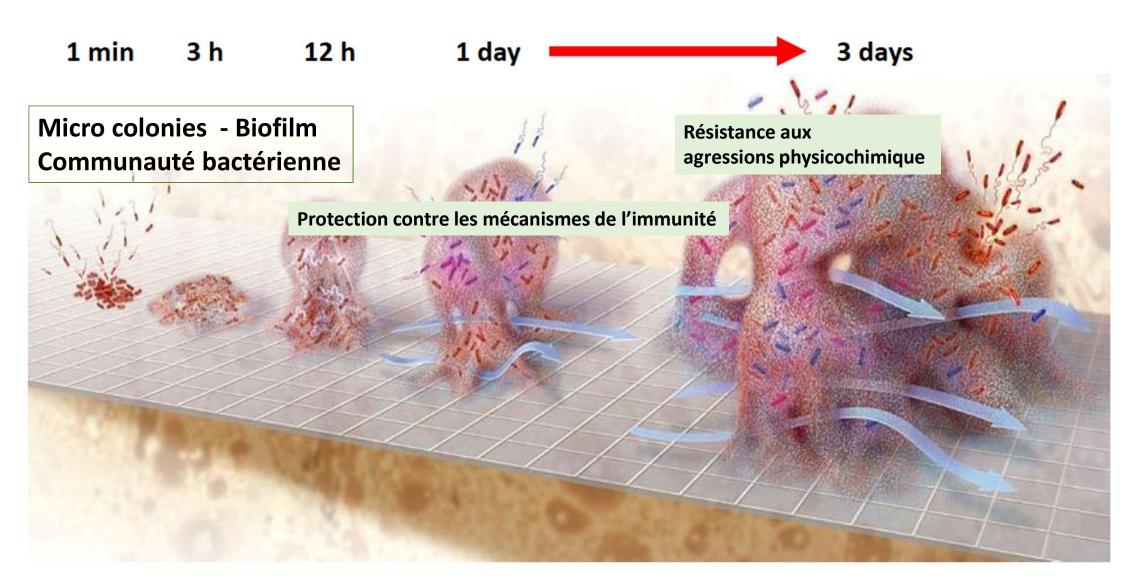
- Apporte un environnement stable pour la pousse bactérienne
- Favorise les échanges entre bactéries
- Stabilise la colonie bactérienne à la surface du substrat



D'après Keith Kasnot, Scientific American 2001

Biofilm rôle

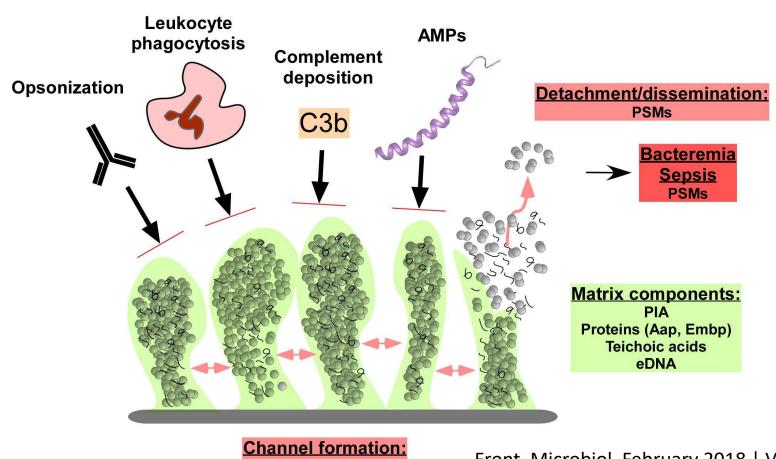
- Apporte un environnement stable pour la pousse bactérienne
- Favorise les échanges entre bactéries
- Stabilise la colonie bactérienne à la surface du substrat
- Protège d'une grande variété d'agressions possibles:
 - Exposition aux UV
 - toxicité du métal
 - exposition acide
 - Déshydratation
 - substances toxiques



D'après Keith Kasnot, Scientific American 2001

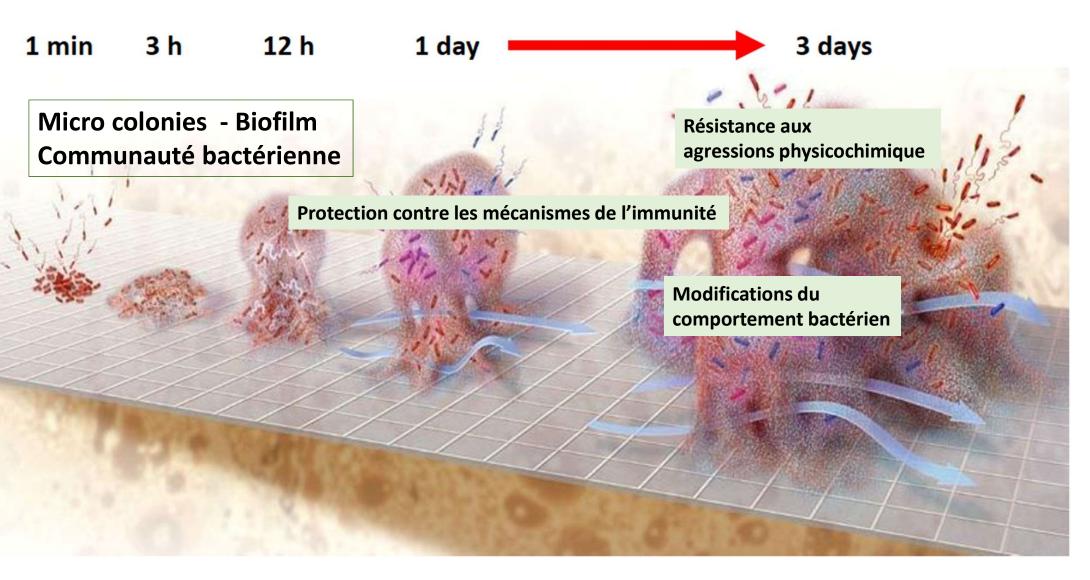
Biofilm et immunologie

BIOFILM INHIBITS:



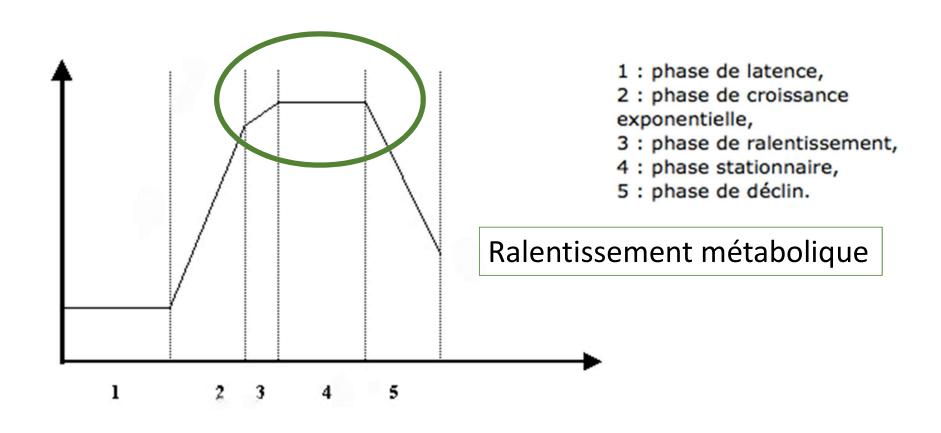
PSMs

Front. Microbiol. February 2018 | Volume 9 | Article 359



D'après Keith Kasnot, Scientific American 2001

Biofilm modification du comportement bactérien. Phase stationnaire de croissance



Biofilm modification du comportement bactérien. Phase stationnaire de croissance

Table 2. Minimum bactericidal concentration (MBC) of Staphylococcus epidermidis B3972.

	Phases of growth (Fold		
Antibiotic	Logarithmic	Stationary	increase	
Vancomycin	4	50	12.5	
Daptomycin	2	12.5	6	
Teicoplanin	4	12.5	3	
Ciprofloxacin	0.5	100	200	
Rifampin	0.06	0.15	2.5	
Netilmicin	8	400	50	

A. F. Widmer, The Journal of Infectious Diseases 1990;162:96-102

Killing of Nongrowing and Adherent *Escherichia coli* Determines Drug Efficacy in Device-Related Infections

ANTIMICROBIAL AGENTS AND CHEMOTHERAPY, Apr. 1991, p. 741–746

ANDREAS F. WIDMER, 1† ADRIAN WIESTNER, 1 RENO FREI, 2 AND WERNER ZIMMERLI 1*

TABLE 4. MBC for E. coli ATCC 25922 in the logarithmic and stationary phases of growth

D	MBC (µg/ml) in:		
Drug	Log phase	Stationary phase ^a	
Co-trimoxazole (TMP/SMX) ^b	0.06/1.2	80/1,600	
Aztreonam	0.125	100	
Fleroxacin	0.07	4	
Ciprofloxacin	0.02	0.02	
Ciprolioxacin	0.02	0.02	

[&]quot; The MBC was determined with bacteria in the stationary phase of growth (see text).

b Trimethoprim-sulfamethoxazole (TMP/SMX) was tested in vitro with a mixture of 1:20.

Staphylococcus epidermidis B3972 Producteur de Slime.

Croissance Logarithmique

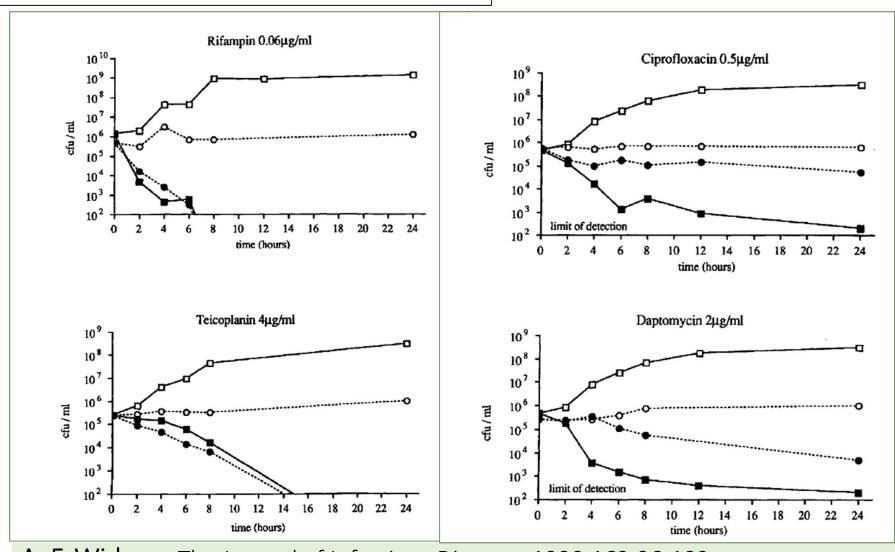
ATB

Contrôle

Croissance stationnaire

ATB

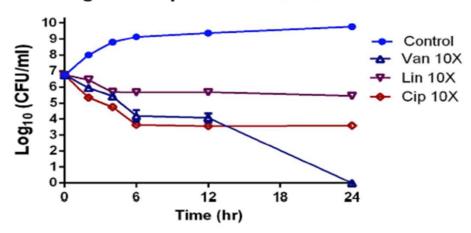
Contrôle



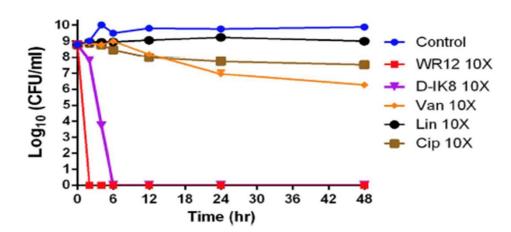
A. F. Widmer, The Journal of Infectious Diseases 1990;162:96-102

Biofilm modification du comportement bactérien. Phase stationnaire de croissance

b-Logarithmic phase of MRSA USA300



d- Stationary phase of MRSA USA300



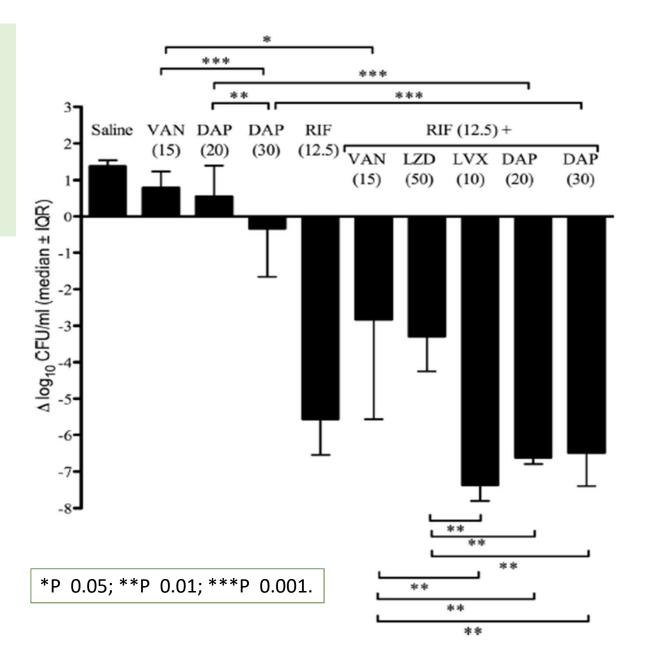
Nature /scientific report 2016 DOI: 10.1038/srep29707

Infection de matériel étranger Efficacité des antibiotiques. Modèles animaux

Micro- organismes	antibiotiques	CMB (phase log)	% guérison	Auteur
S. epidermidis	vancomycine ciprofloxacine rifampicine	4 0.5 0.06	17 0 100	Widmer, JID 1990
S. aureus	vancomycine	2.5	0	Frei,
	ciprofloxacine	0.62	17	ICAAC
	rifampicine	0.12	50	1990
E. coli	cotrimoxazole	0.06	0	Widmer,
	ciprofloxacine	0.02	92	AAC 1991

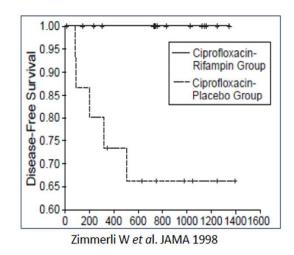
Efficacy of Daptomycin in Implant-Associated Infection Due to Methicillin-Resistant Staphylococcus aureus: Importance of Combination with Rifampicin

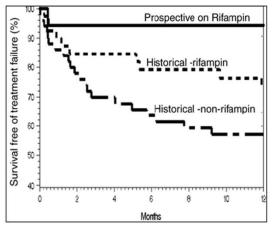
Antimicrob. Agents Chemother, July 2009, p. 2719–2724



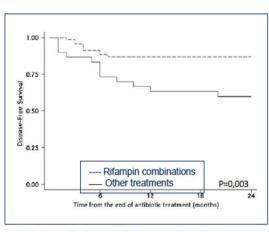
Infections de Prothèses articulaires à S. aureus. Efficacité selon les ATB

Combinaisons de Rifampicine dans les IPOA à staphylocoques (S. aureus et SCN)

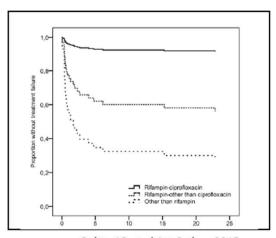




El Helou OC et al. Eur J Clin Microbiol Infect Dis. 2010

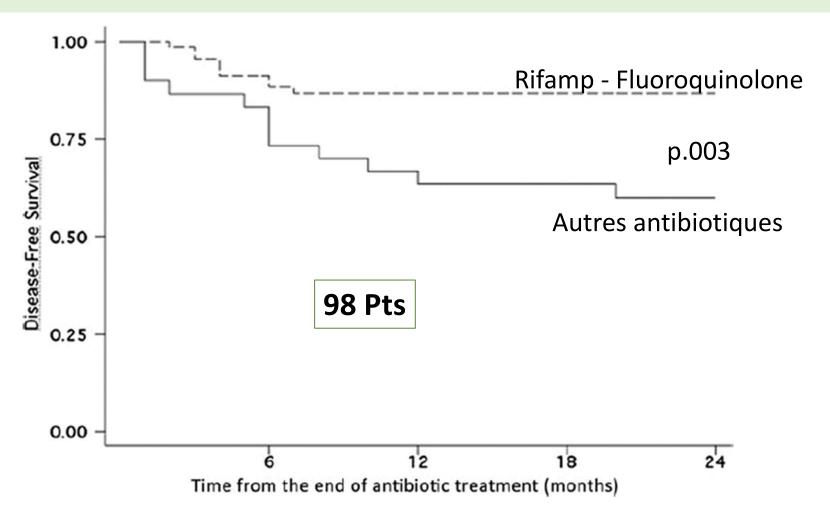


Senneville E et al. Clin Infect Dis 2011



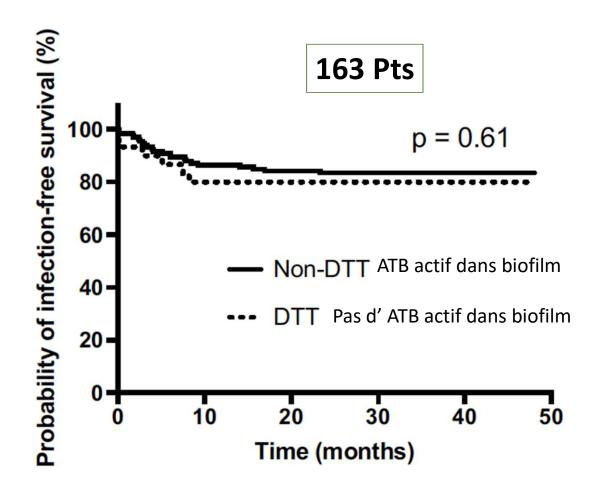
Puhto AP et al. Int Orthop 2015

Infections de Prothèses articulaires à S. aureus. Efficacité selon les ATB



E Senneville Clinical Infectious Diseases 2011;53(4):334–340

Infections de Prothèses articulaires. Efficacité selon les ATB



Archives of Orthopaedic and Trauma Surgery oct 2017 doi10.1007/s00402-018-2886-0

Infections de Prothèses articulaires. Efficacité selon les ATB

Table 2 Treatment and outcome characteristics of all patients		Pas d' ATB actif dans biofilm	ATB actif dans biofilm	
Variable	All patients $(n=163)$	DTT PJI group $(n=30)$	Non-DTT PJI group $(n=133)$	p value
Treatment				
Time until reimplantation (days)	63.9 ± 34.5	89.4 ± 50.5	58.1 ± 26.9	< 0.001
CRP prior to reimplantation (mg/l)	11.9 ± 13.1	10 ± 11.8	12.3 ± 13.4	0.164
Total duration of antimicrobial therapy (days)	123.3 ± 57.7	150.8 ± 74.7	117.4 ± 51.7	0.003
Duration of i.v. antimicrobial therapy (days) ^a	32.8 ± 20.2	50.4 ± 31	29 ± 14.7	< 0.001
Duration of oral antimicrobial therapy (days) ^a	84.1 ± 32.2	90.1 ± 47.3	82.8 ± 28.2	0.553
No. of revisions during interval	1.7 ± 1.3	2 ± 1.6	1.5 ± 1	0.324
Duration of hospital stay (days)	31.2 ± 15.9	44.5 ± 26.5	28.2 ± 10.4	< 0.001

Archives of Orthopaedic and Trauma Surgery oct 2017 doi10.1007/s00402-018-2886-0

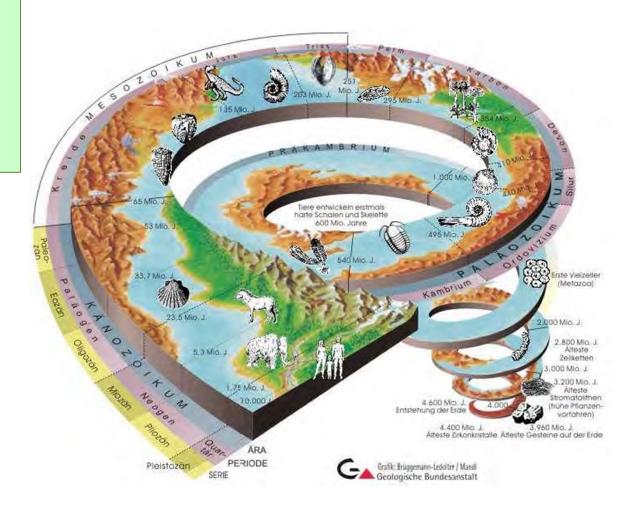
Les bactéries productrices de slime

- Staphylococcus aureus
- Staphylococcus epidermidis
- Streptococcus sp (non groupables)
- Enterococcus sp
- Pseudomonas aeruginosa
- E coli, Enterobacter sp, Proteus sp
- Anaérobies : bacteroides sp, corynebactéries

1/ Adhérence et organisations bactériennes

2/ Variants microcolonies

3/ Persistance intra cellulaire



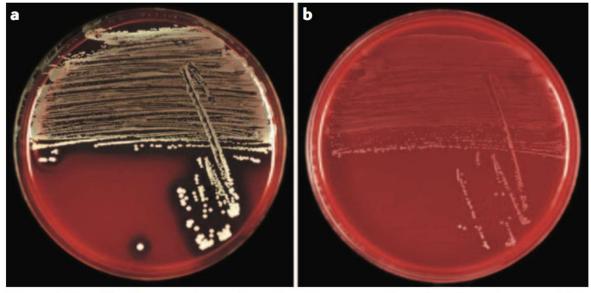


Figure 1 | **Small colony variants.** Columbia blood-agar plates that show the normal (a) and the small colony variant (b) phenotype of *Staphylococcus aureus* are shown.

Nat Rev Microbiol 2006(4)295

Phénomène d'adaptation auxotrophique

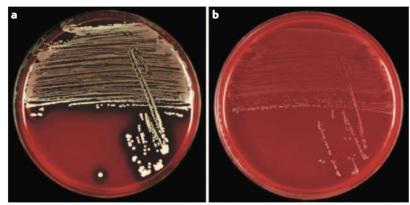


Figure 1 | **Small colony variants.** Columbia blood-agar plates that show the normal (a) and the small colony variant (b) phenotype of *Staphylococcus aureus* are shown.

- > Déficit en transport d'électron
 - Altération de synthèse de medianone & hémine
 - Déficit en biosynthèse de thymidine

Nat Rev Microbiol 2006(4)295

Modifications phénotypiques

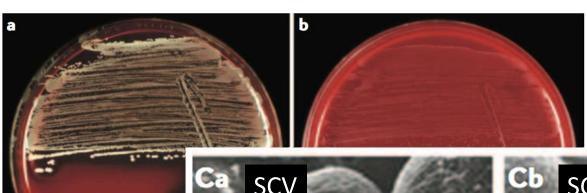
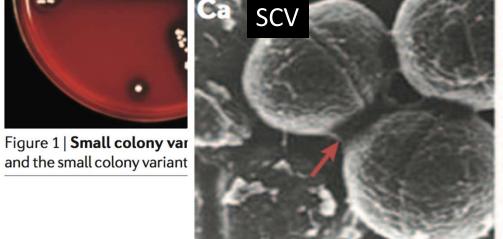
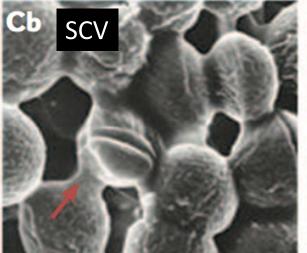
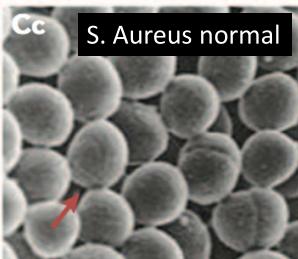


Figure 5 | Altered cellular morphology of haemin-auxotrophic small colony







Nat Rev Microbiol 2006(4)295

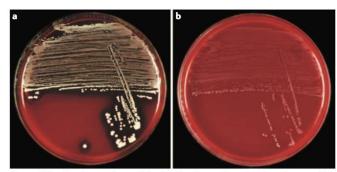


Figure 1 | **Small colony variants.** Columbia blood-agar plates that show the normal (a) and the small colony variant (b) phenotype of *Staphylococcus aureus* are shown.

Petites colonies

Pousse lente

- 🔪 Activité oxydatives / métaboliques
- ∠ Capacités d'adhérence
- > Production de certains facteurs de virulence

Nat Rev Microbiol 2006(4)295

Small Colony Variants

Petites colonies

Pousse lente

Activité oxydatives / métaboliques

Capacités d'adhérence

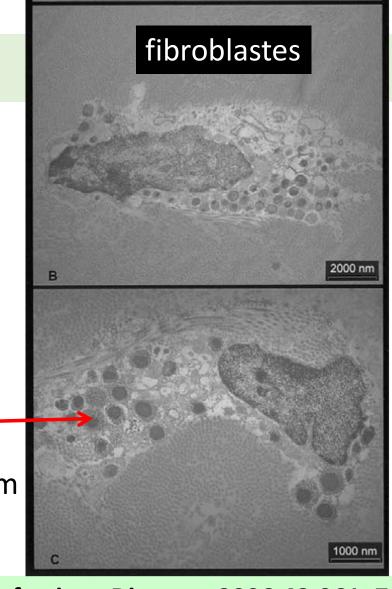
Production de certains facteur

de virulence

Uprégulation des gènes de persistance

Facilitation de la persistance intracellulaire

Uprégulation des gène de formation du biofilm



Sendi Clinical Infectious Diseases 2006;43:961-7

SCV

Small Colony Variants

Petites colonies
Pousse lente
Activité oxydatives / métaboliques
Capacités d'adhérence
Production de certains facteur
de virulence

Uprégulation des gènes de persistance

Uprégulation des gène de formation du biofilm

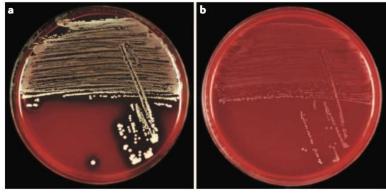
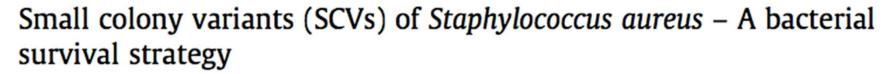


Figure 1 | **Small colony variants.** Columbia blood-agar plates that show the normal (a) and the small colony variant (b) phenotype of *Staphylococcus aureus* are shown.



Barbara C. Kahl*

Small Colony Variants

Persistent and Relapsing Infections Associated with Small-Colony Variants of Staphylococcus aureus

Richard A. Proctor, Petra van Langevelde, Mar Kristjansson, Joel N. Maslow, and Robert D. Arbeit From the Departments of Medicine and Medical Microbiology and Immunology, University of Wisconsin-Madison Medical School, Madison, Wisconsin; and the Department of Medicine, Veterans Affairs Medical Center, and the Departments of Medicine and Microbiology, Boston University School of Medicine, Boston, Massachusetts

Table 1. Clinical characteristics of patients with persistent and relapsing infection associated with S. aureus SCVs.

Case no.	Type(s) of infection (site)	Interval between episodes of infection	Duration of clinically persistent infection despite treatment	Change in treatment regimen closely associated with clinical response
1	Osteomyelitis (femur)	54 y	6 d	Substitution of clindamycin (iv) for vancomycin
2	Osteomyelitis (femur)	29 y	>4 w	*
3	Sinusitis (maxillary sinus)	4 mo	>3 w	*
4	Osteomyelitis, septic arthritis (hip prosthesis)	6 mo	8 w	Surgery (removal of prosthesis)
5	Muscle abscesses, septic arthritis (thighs, hips)	4 y [†]	12 w	Addition of vitamin K and TMP-SMZ to vancomycin, rifampin

^{*} There was a very delayed response to continued conventional treatment with parenteral β-lactam antibiotics (see text for details).

[†] Isolate from initial episode of infection was not identified as S. aureus but was phenotypically consistent with SCVs (see text for de CID 1995;20 (January)

Small Colony Variants relation avec les antibiotiques

Résistance par mécanisme spécifique décrite pour fluoroquinolones, aminosides, Co trimoxazole

Induction de formation de SCV par exposition à Co-trimoxazole, fluoroquinolone

Small Colony Variants Sensibilité Antibiotiques

Clinical Characteristics and Outcomes of Prosthetic Joint Infection Caused by Small Colony Variant Staphylococci

TABLE 3 Antimicrobial susceptibility and auxotrophy testing results for isolates recovered from subjects with SCVs

Susceptibility or auxotrophy result	20 (47.6) 21 (6 36 (85.7) 30 (8 37 (88.1) 33 (9	status:
	$\overline{\text{Yes } (n=42)}$	No $(n = 34)$
Antimicrobial susceptibility testing		
Oxacillin susceptiblea	20 (47.6)	21 (61.8)
Gentamicin susceptible	36 (85.7)	30 (88.2)
Rifampin susceptible	37 (88.1)	33 (97.1)
Minocycline susceptible	42 (100)	33 (97.1)
Vancomycin susceptible	42 (100)	34 (100)
Trimethoprim-sulfamethoxazole susceptible ^b	34 (85.0)	31 (91.2)

Tande et al Mbio 2014

Small Colony Variants Sensibilité Antibiotiques

AAC 2018 6	2 Issue 4	MIC data (µ		ureus Paires		& souch	ne parent		
			NP			SCV			
	Antimicrobial agent	MIC ₅₀	MIC ₉₀	Range	MIC ₅₀	MIC ₉₀	Range		
	β -Lactams								
CC	Benzylpenicillin	0.094	2	0.023-24	0.125	1.5	0.016-4		
	Ampicillin	0.25	3	0.094-12	0.25	1.5 (2)	0.023-4 (0.023-6)		
	Ampicillin-sulbactam	0.19 (0.25)	1	0.094-1.5	0.19	0.75	0.016-0.75 (0.016-1)		
	Piperacillin	1.5	4	0.75-24 (0.75-32)	1.5	3	0.25-4 (0.25-12)		
	Piperacillin-tazobactam	1	2	0.75-3	0.5	1	0.064-2		
	Oxacillin	0.38	0.75	0.19-1	0.125	0.5	0.023-0.5		
	Cefoxitin	2	3	1–3	1	3	0.25-4		
	Cephalothin	0.38	0.5	0.19-0.75	0.125	0.38	0.016-0.5		
	Cefuroxime	0.75	0.75	0.38-1.5	0.25	0.75	0.064-0.75		
	Ceftriaxone	3	4	2-6	2	4	0.38-4		
	Cefepime	3	3	1.5-3 (1.5-4)	1.5	3	0.38-3		
	Imipenem	0.064	0.094	0.047-0.094	0.023	0.064	0.012-0.064		

Small Colony Variants Sensibilité Antibiotiques

AAC 2018 62 Issue 4 10 souches S.aureuscPaires de SCVc& souche parent

MIC data (μg/ml)^a

	NP			SCV		
Antimicrobial agent	MIC ₅₀	MIC ₉₀	Range	MIC ₅₀	MIC ₉₀	Range
Non-β-lactams						
Vancomycin	1	1.5	0.75-1.5	1	1.5	0.75-2
Daptomycin	0.19	0.25	0.094-0.38	0.094 (0.19)	0.38	0.094-0.5
Moxifloxacin	0.064	0.064	0.032-0.064	0.047	0.19	0.023-0.19
Clarithromycin	0.38	0.75	0.094-256	0.19	1.5	0.023-256
Clindamycin	0.094	0.125	0.064-0.125	0.064	0.19	0.016-0.38
Linezolid	0.5 (0.75)	1 (1.5)	0.25-1 (0.25-1.5)	0.38	1.5	0.094-1.5
Rifampin	0.012	0.016	0.006-0.016	0.008	0.012	0.003-0.016
Fosfomycin	4	12	1.5-16	4	12	0.064-16
Quinupristin-dalfopristin	0.5	0.5 (0.75)	0.25-0.75	0.38	0.5	0.047-0.75
Tigecycline	0.38	0.5	0.19-0.75	0.38	0.5 (0.75)	0.094-0.75
Trimethoprim- sulfamethoxazole	0.032	0.047	0.023-0.19	0.032	0.094	0.003-0.125
Gentamicin	0.38	24	0.125-256	0.125 (0.25)	4	0.064-256
Mupirocin	0.094	0.19 (0.38)	0.094-0.19 (0.094-0.38)	0.094	0.19	0.064-0.19

Small Colony Variants conséquences IOA

Clinical Characteristics and Outcomes of Prosthetic Joint Infection Caused by Small Colony Variant Staphylococci

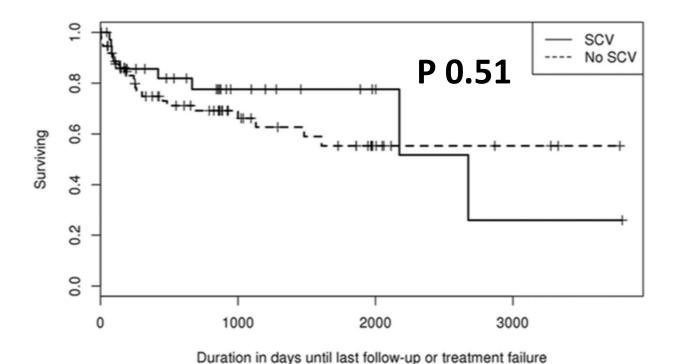
Aaron J. Tande, a,b Douglas R. Osmon, Kerryl E. Greenwood-Quaintance, Tad M. Mabry, Arlen D. Hanssen, Robin Patela,b,d

Tande et al Mbio 2014

TABLE 1 Medical and orthopedic sur	gical history and PII presentation			
Characteristic ^d	SCV ^b			
	Yes (n = 38)	No (n = 75)	P value	
Demographic factors Age in yr, median (range) Female sex Diabetes mellitus Rheumatoid arthritis by ACR criteria Rheumatoid or inflammatory arthritis CKD	64 (25–85) 16 (42.1) 10 (26.3) 1 (2.6)	63 (36–84) 31 (41.3) 20 (26.7) 5 (6.7) 11 (14.7) 6 (8.0)	0.49 1 0.97 0.66 0.38 0.72	
Orthopedic history				
Joint age in days, median (range) Prior arthroplasty revision	1,295 (216–13,712) 32 (84.2)	646 (23–11,883) 52 (70.3)		0.007 0.17
Time since last surgery in days,	743 (31–10,030)			<0.0001
median (range) Cemented arthroplasty	33 (86.8)	60 (80.0)	0.44	
Duration of PJI symptoms in days,	491 (14–2,306)	165 (2–1,656)		0.0003
median (range)				
median (range) Prior surgery for this PII	23 (60.5)	28 (37.3)	0.03	
Receiving 120 or more days of	16 (42.1)	17 (22.7)		0.048
antibiotics in prior 6 mo				
median (range) Serum ESR in mm/h, median (range)	46 (5–111)	43 (3–123)	0.54	
Serum CRP in mg/liter,	23 (5–222)	44 (3–269)	0.2	
median (range) Preoperative SF aspirate SF WBC in cells/ μ l, median (range)	26 (68.4) 28,574 (8,175–155,000)	51 (68.0) 44,275 (629–1,071,472)	0.13	
SF neutrophil %, median (range)	88 (79–98)	^{91 (51–100)} Tand	e et al Mbio	2014

Clinical Characteristics and Outcomes of Prosthetic Joint Infection Caused by Small Colony Variant Staphylococci

Aaron J. Tande, a,b Douglas R. Osmon, Kerryl E. Greenwood-Quaintance, Tad M. Mabry, Arlen D. Hanssen, Robin Patela,b,d



Remplacement prothétique en 2 temps 100 % des patients

Tande et al Mbio 2014

Small Colony Variants : bactéries pour lesquelles des phénotypes SCV ont été décrits

Staphylococcus aureus

Staphylococcus coagulase négative

Pseudomonas aeruginosa

E. coli,

N. Gonorrhoeae

S. Typhimurium Shigella

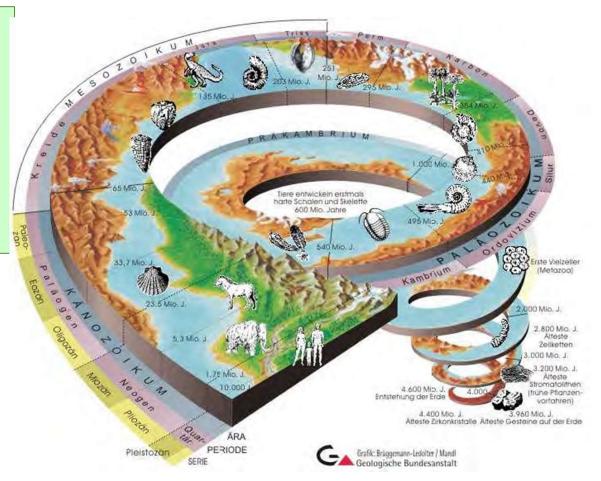
Proteus sp., Klebsiella pneumoniae, Providencia stuartii,

Enterobacter cloacae, S. marcescens, Citrobacter freundii

1/ Adhérence et organisations bactériennes

2/ Variants microcolonies

3/ Persistance intra cellulaire



Microcolonies formant biofilm identifiées :

- 3.3–3.4 milliard d'années South African Kornberg formation
- 3.2-milliard d'années deep-sea hydrothermal rocks of the Pilbara Craton, Australia

Persistance intracellulaire

Internalisation dans les cellules non phagocytaires



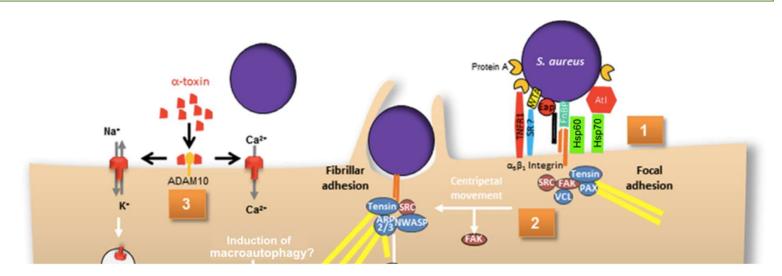
Persistance intracellulaire



Échappement du compartiment intra cellulaire

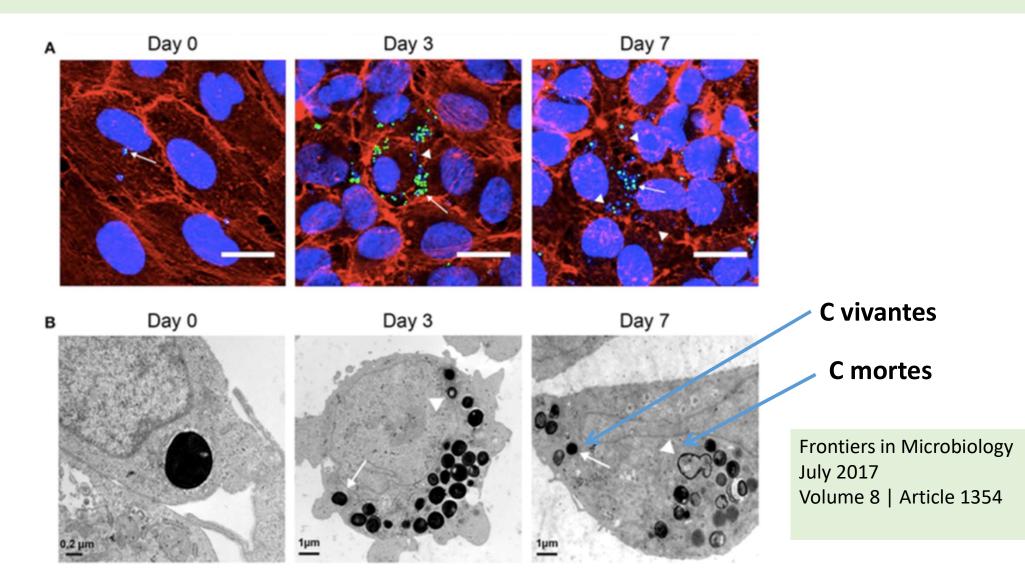
Persistance intracellulaire

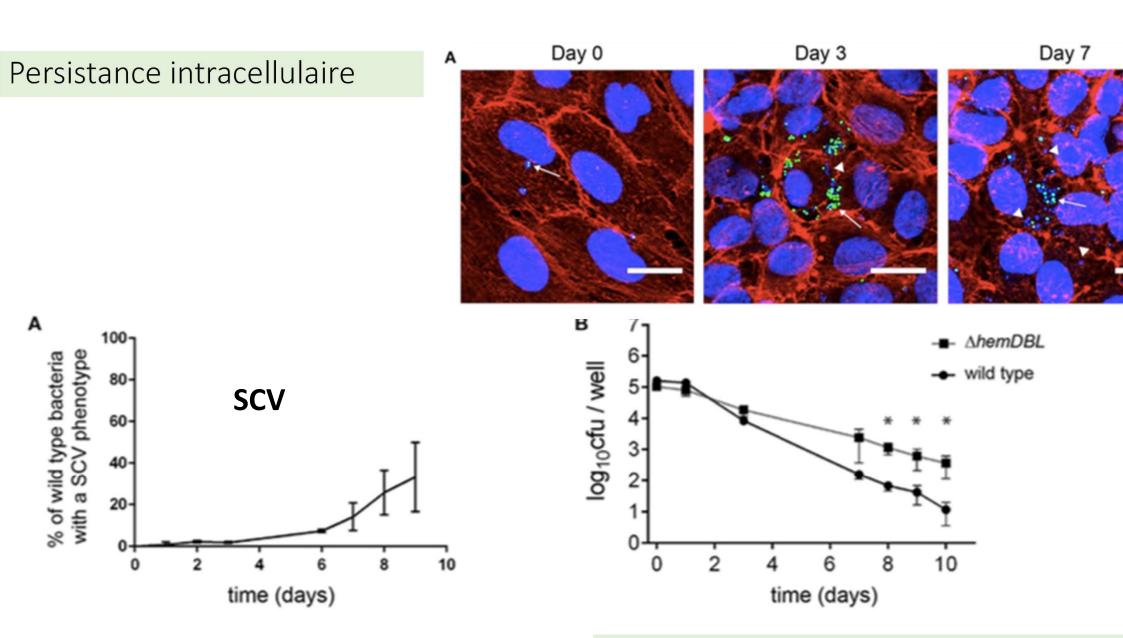
Internalisation dans les cellules non phagocytaires



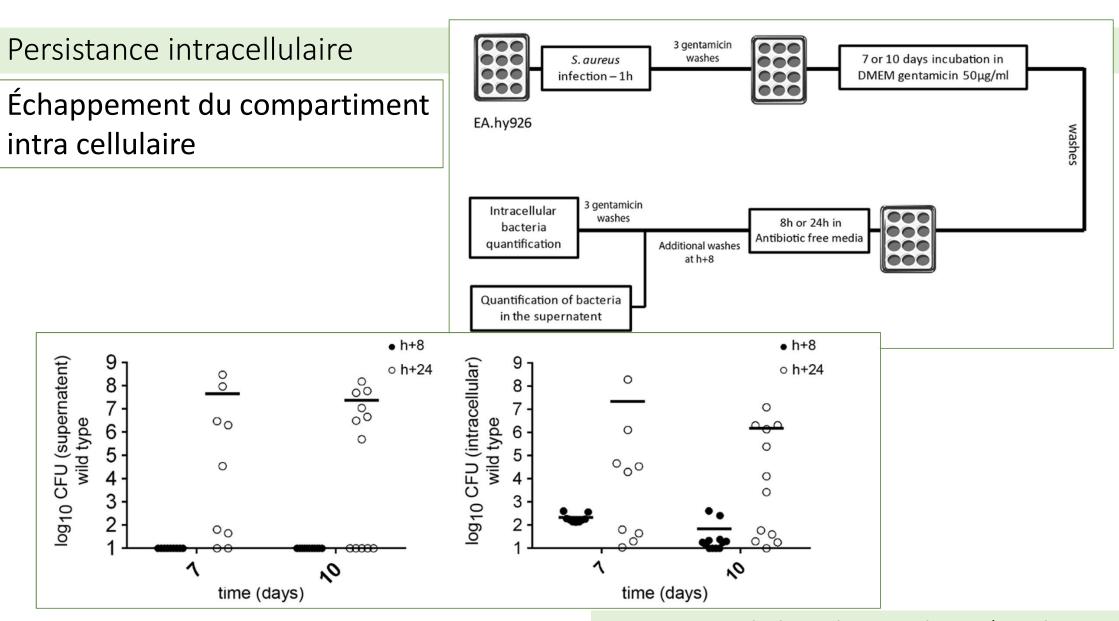
Fibroblastes Cellules endothéliales Ostéoblastes kératinocytes

Persistance intracellulaire





Frontiers in Microbiology July 2017 Volume 8 | Article 1354



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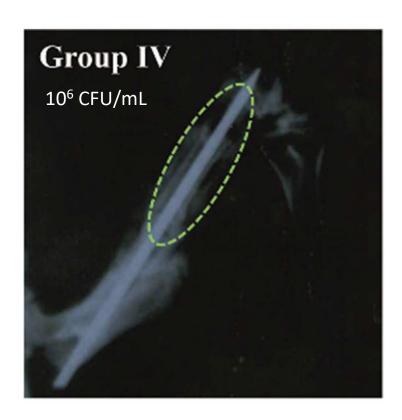
INTRA-CELLULAR STAPHYLOCOCCUS AUREUS ALONE CAUSES INFECTION IN VIVO

Therwa Hamza^{1,2}, Matthew Dietz¹, Danh Pham¹, Nina Clovis¹, Suzanne Danley¹ and Bingyun Li^{1,2,3,4,*}

Rat bone Inoculation 10⁶ osteoblasts infected with 10⁶ CFU intra- cellular *S. aureus*



Day 21

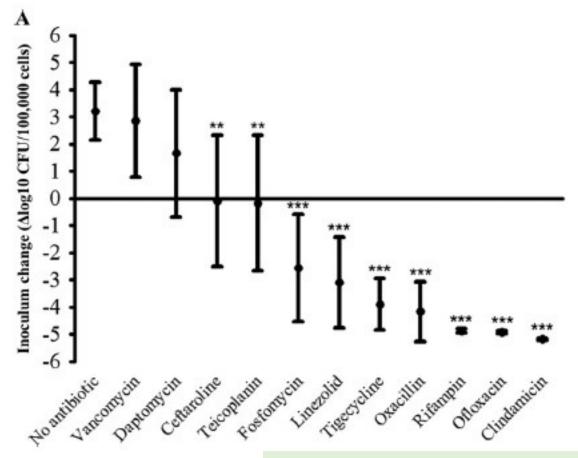


Persistance intracellulaire Antibiotiques actifs



Antimicrobial Activity against Intraosteoblastic Staphylococcus aureus

Inoculum intra ostéoblastique



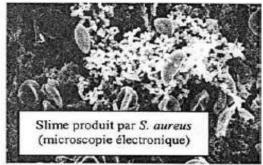
F. Valour Antimicrob Agents Chemother 201559:2029 –2036.

Les altérations du métabolisme bactérien au voisinage des matériaux infectés responsable d'un phénomène de tolérance aux antibiotiques, même bactéricides (small colony variant)

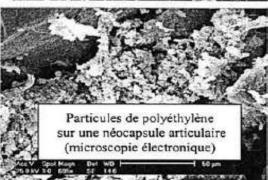
Le slime (substance polysacccharidique extracellulaire)

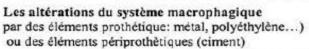
et le biofilm bactérien semblant responsable de :

- une adhésion bactérienne plus forte,
- une virulence particulière et
- une moindre sensibilité aux antibiotiques











Infections de prothèse articulaire

Prosthetic joint infections

L. Bernard *

Les réservoirs de germes inaccessibles aux antibiotiques (ciment, granulome...)



Médecine et maladies infectieuses 33 (2003) 231-239

1/ Adhérence et organisations bactériennes

2/ Variants microcolonies

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Merci de votre attention

