

What is the Future of Microbiologic Diagnosis?



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Déclaration d'intérêts

- **Contracted Research: ContraFect, TenNor Therapeutics Limited, and BioFire**
- **Consultant: Curetis, PathoQuest, Selux Diagnostics, 1928 Diagnostics, PhAST, Torus Biosystems, Day Zero Diagnostics, Mammoth Biosciences, CARB-X, Qvella, Netflix**
- **Mayo Clinic and I have a relationship with Adaptive Phage Therapeutics and Pathogenomix**
- **Patents: *Bordetella pertussis/parapertussis* PCR; device/method for sonication; anti-biofilm substance**

Outline

- **The challenge**
- **Technology revolution**
 - Proteomics
 - NAATs & sequencing-based diagnostics
 - Other diagnostic types (imaging, metabolomic)
 - Point of care diagnostics
 - Advanced host response assessment for infectious diseases
- **How to move to the future**
 - Demonstration of clinical utility
- **The deliverables**
 - Improved health
 - Changes to healthcare delivery
 - Better understanding of infectious diseases and their mimics

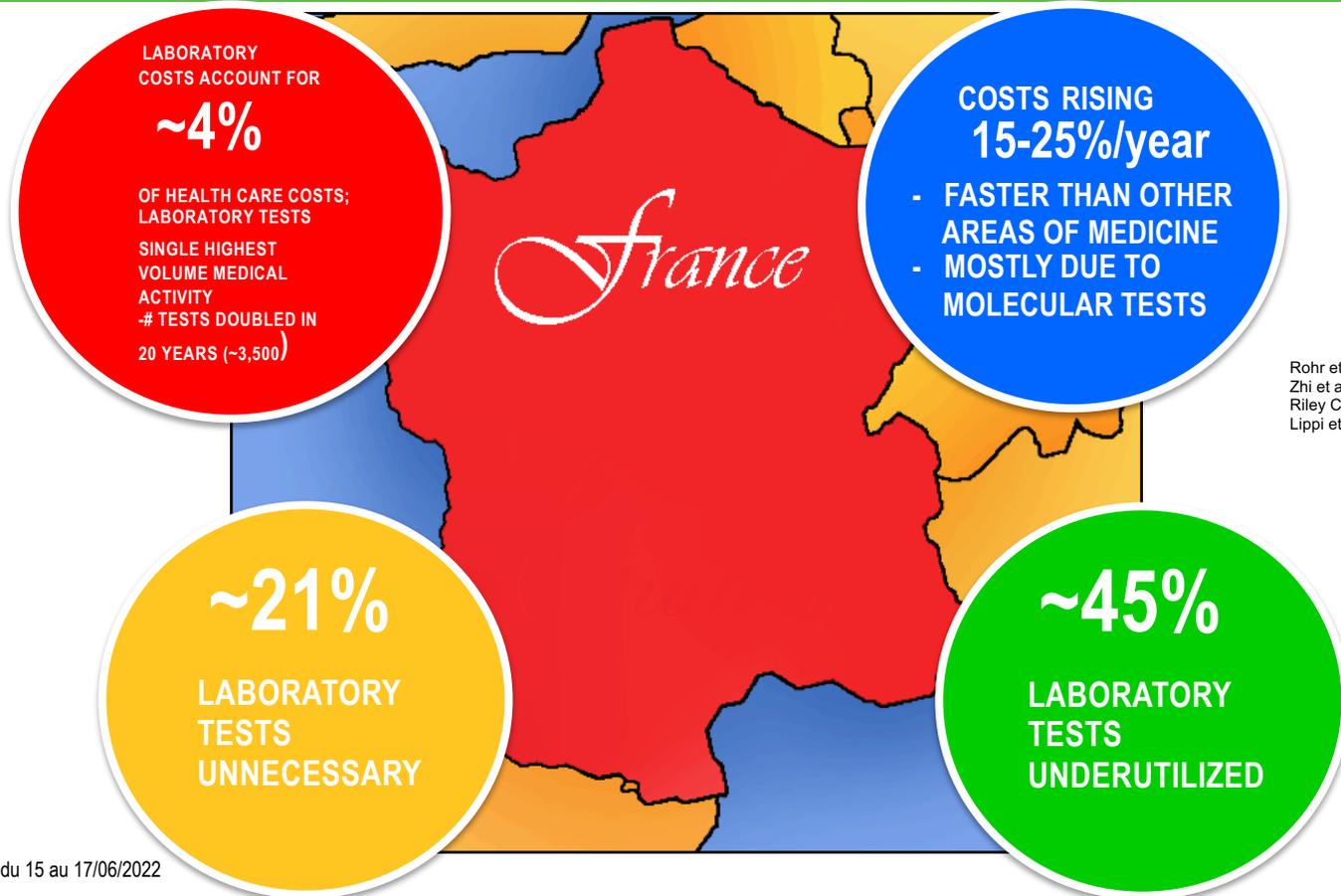
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The Challenge

- **The need:**
 - Antimicrobial resistance
 - Emerging infectious diseases
 - Availability of better therapeutics
- **The barriers:**
 - Infectious diseases management predicated on “culture of empiricism”
 - Inappropriate test utilization
 - Lack of understanding of value of advanced diagnostics
 - Cost of advanced diagnostics

70% Clinical Decisions Substantially Based on Results of Diagnostic Tests

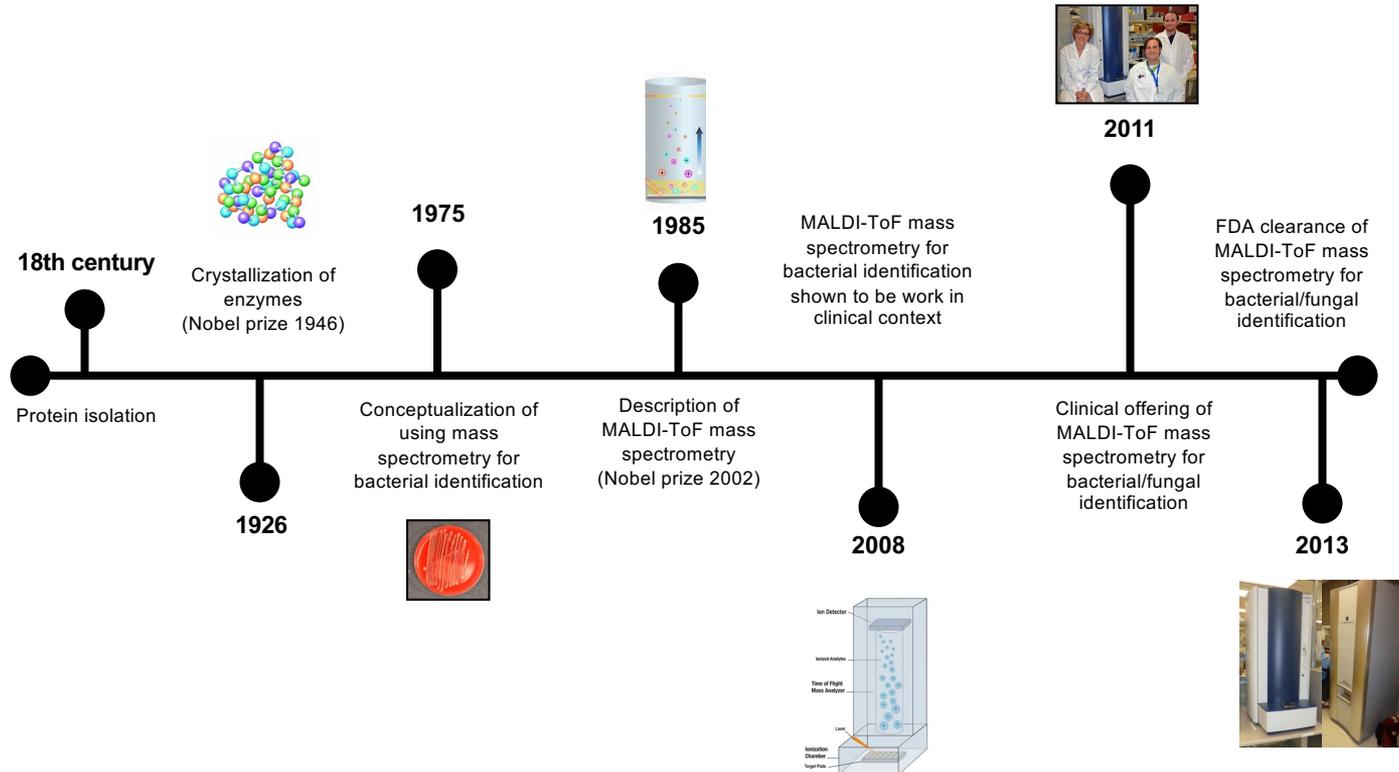


Rohr et al. PLoS One 2016;11:e0149856
Zhi et al. PLoS One. 2013;8(11):e78962
Riley Clin Microbiol Newslett 2017;39:69-73
Lippi et al. Ann Transl Med 2017;5(4):82

Outline

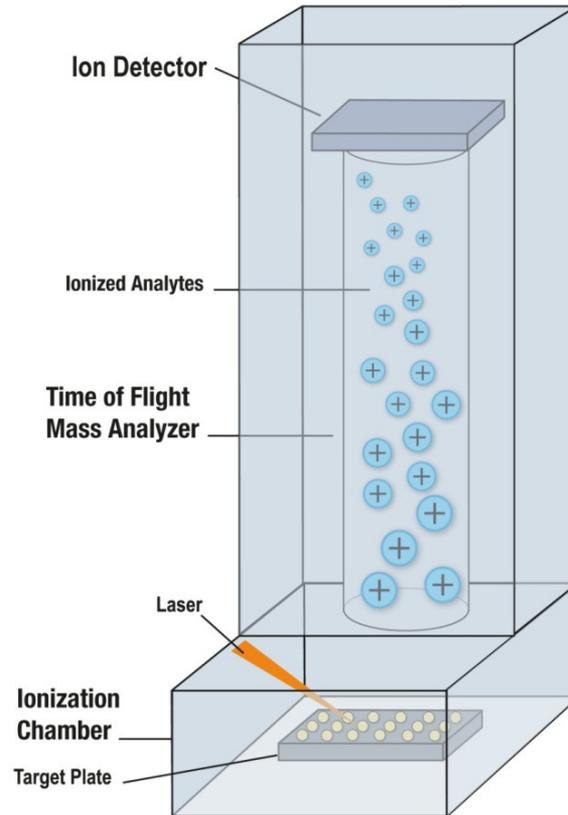
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Technology Revolution Proteomics



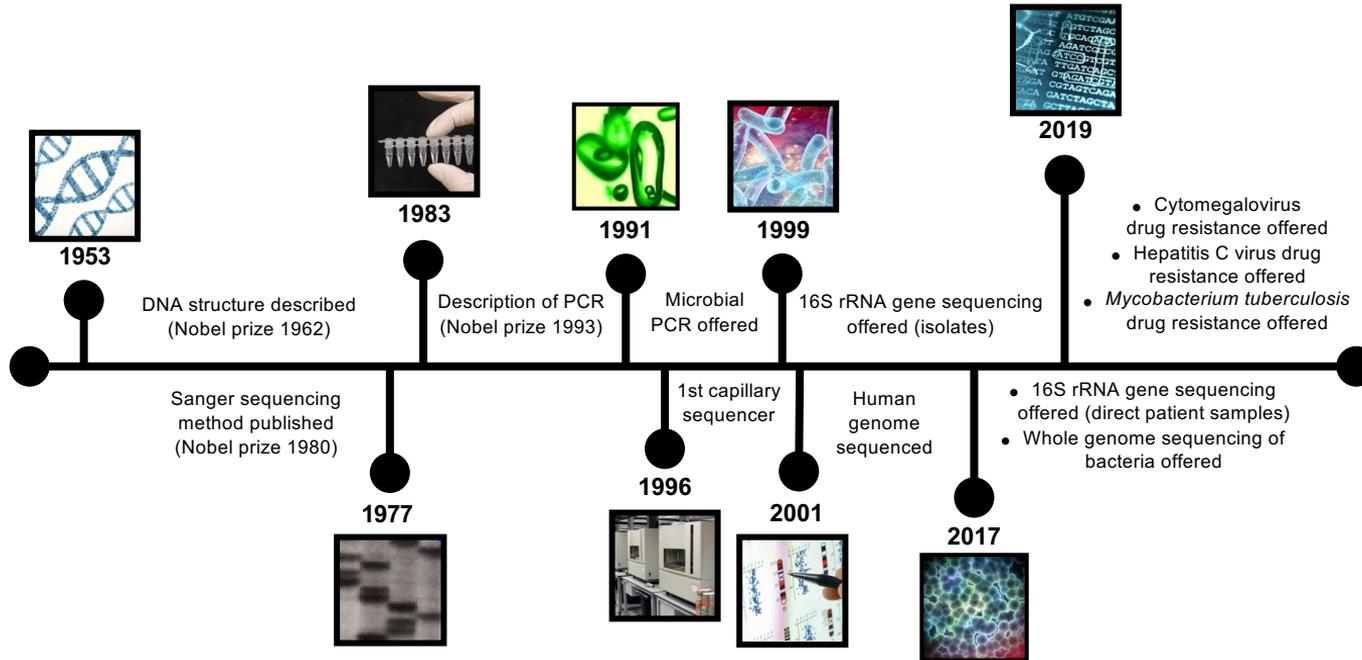
Technology Revolution

MALDI ToF Mass Spectrometry



Technology Revolution

NAAT and Sequencing-Based Diagnostics



Technology Revolution: Microbial Real-Time PCR Assays Developed @ Mayo Clinic, 2000–2020

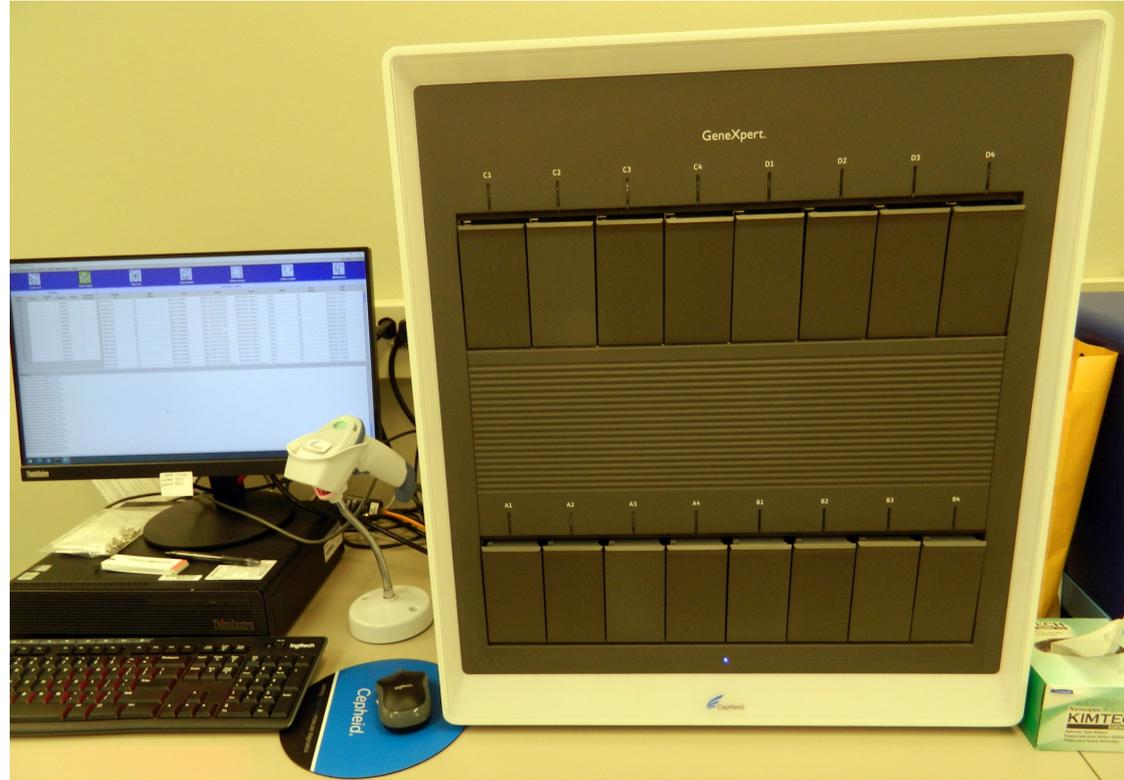
| | |
|------|---|
| 2000 | Herpes simplex virus* |
| 2001 | <i>Bacillus anthracis</i> |
| | Cytomegalovirus* |
| | <i>Toxoplasma gondii</i> * |
| 2002 | <i>Bordetella pertussis/parapertussis</i> * |
| | <i>Streptococcus pyogenes</i> |
| | Enterovirus |
| | Varicella zoster virus* |
| 2003 | <i>Tropheryma whipplei</i> * |
| | <i>Babesia microti</i> * |
| | BK virus* |
| | <i>Ehrlichia/Anaplasma</i> species* |
| | Epstein–Barr virus (qualitative)* |
| | JC virus* |
| | <i>Borrelia burgdorferi</i> * |
| 2004 | <i>Bartonella</i> species* |
| | <i>vanA/vanB</i> * |
| | Parvovirus B19* |
| 2005 | Influenza A/B |
| | <i>Pneumocystis jirovecii</i> |
| | BK virus (quantitative) |
| | Epstein–Barr virus (quantitative) |
| | Human herpesvirus-6 |
| | <i>Plasmodium</i> species |
| | West Nile virus |
| 2007 | <i>Clostridioides difficile</i> |
| | <i>Coccidioides immitis/posadasii</i> |
| | <i>Staphylococcus aureus</i> |
| | <i>Mycobacterium chelonae/abscessus</i> |
| 2008 | Adenovirus |

| | |
|------|---|
| 2009 | <i>Legionella</i> species |
| | <i>bla</i> ^{KPC} |
| | <i>Mycobacterium tuberculosis</i> complex |
| 2010 | <i>Salmonella</i> species |
| | <i>Shigella</i> species |
| | <i>Campylobacter</i> species |
| | <i>Yersinia</i> species |
| | Shiga toxin |
| 2011 | <i>Mycoplasma genitalium</i> |
| | <i>Mycoplasma hominis</i> |
| | <i>Ureaplasma urealyticum/parvum</i> |
| | <i>Histoplasma/Blastomyces</i> species |
| 2012 | <i>Mycobacterium tuberculosis</i> complex species |
| 2013 | <i>Coxiella burnetii</i> |
| | <i>bla</i> ^{NDM} |
| | <i>Mycoplasma pneumoniae</i> |
| 2014 | <i>Babesia</i> species |
| | <i>Borrelia mayonii</i> |
| 2015 | Microsporidia |
| 2016 | <i>Borrelia miyamotoi</i> |
| 2017 | <i>bla</i> ^{OXA48-like} |
| | <i>bla</i> ^{VIM} |
| | <i>Kingella kingae</i> |
| | <i>Acanthamoeba</i> species |
| | Free-living amoeba |
| | Norovirus |
| 2020 | <i>mecA</i> |
| | <i>Helicobacter pylori</i> |
| | <i>Candida auris</i> |
| | SARS coronavirus-2 |

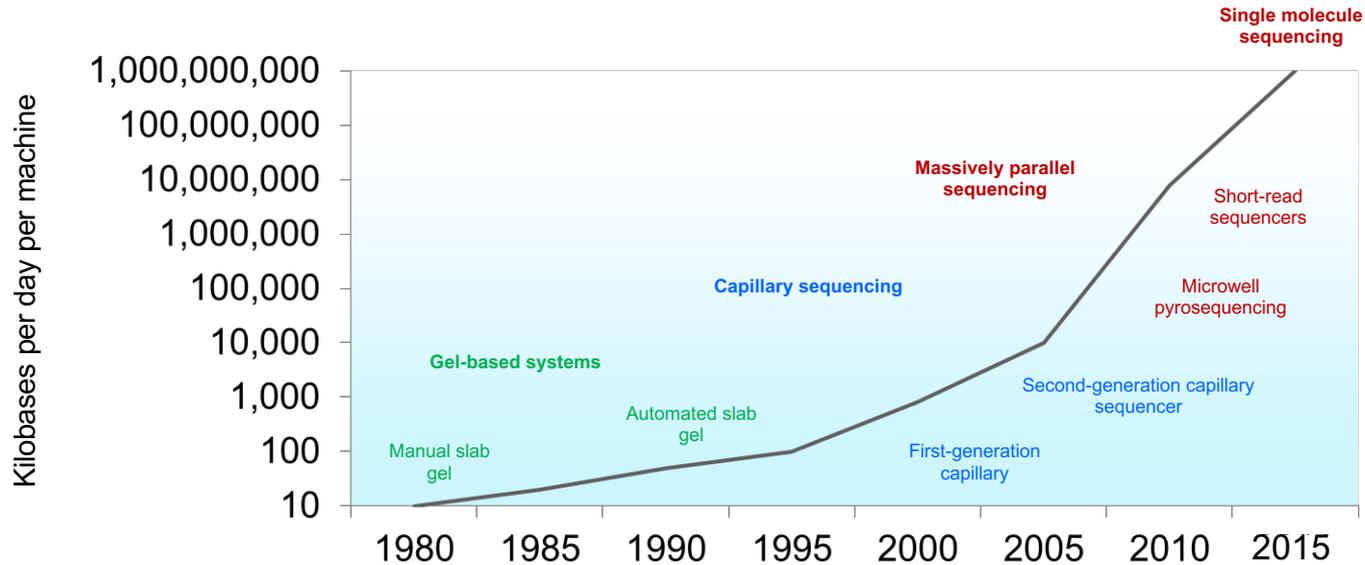
Microorganisms marked by an asterisk (*) were detected with conventional PCR assays with Southern blot detection developed and deployed in the 1990s prior to conversion to real-time PCR assays

Patel. Clin Chem. 2022;68:10-15

Technology Revolution: Multiplex and Rapid Automated NAATS

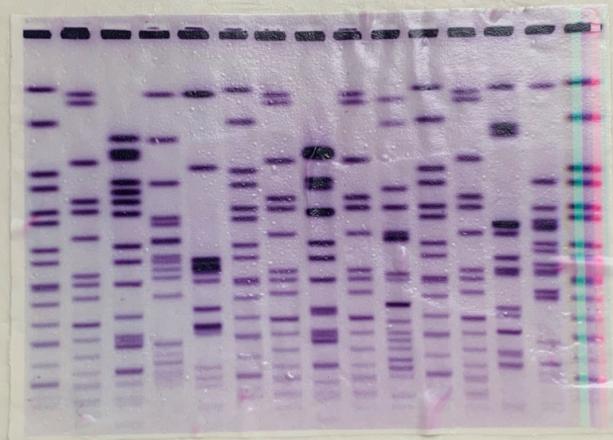


Technology Revolution: Improvements in DNA Sequencing Rates



Based on Stratton et al. Nature 2009;458:719

Farewell PFGE!

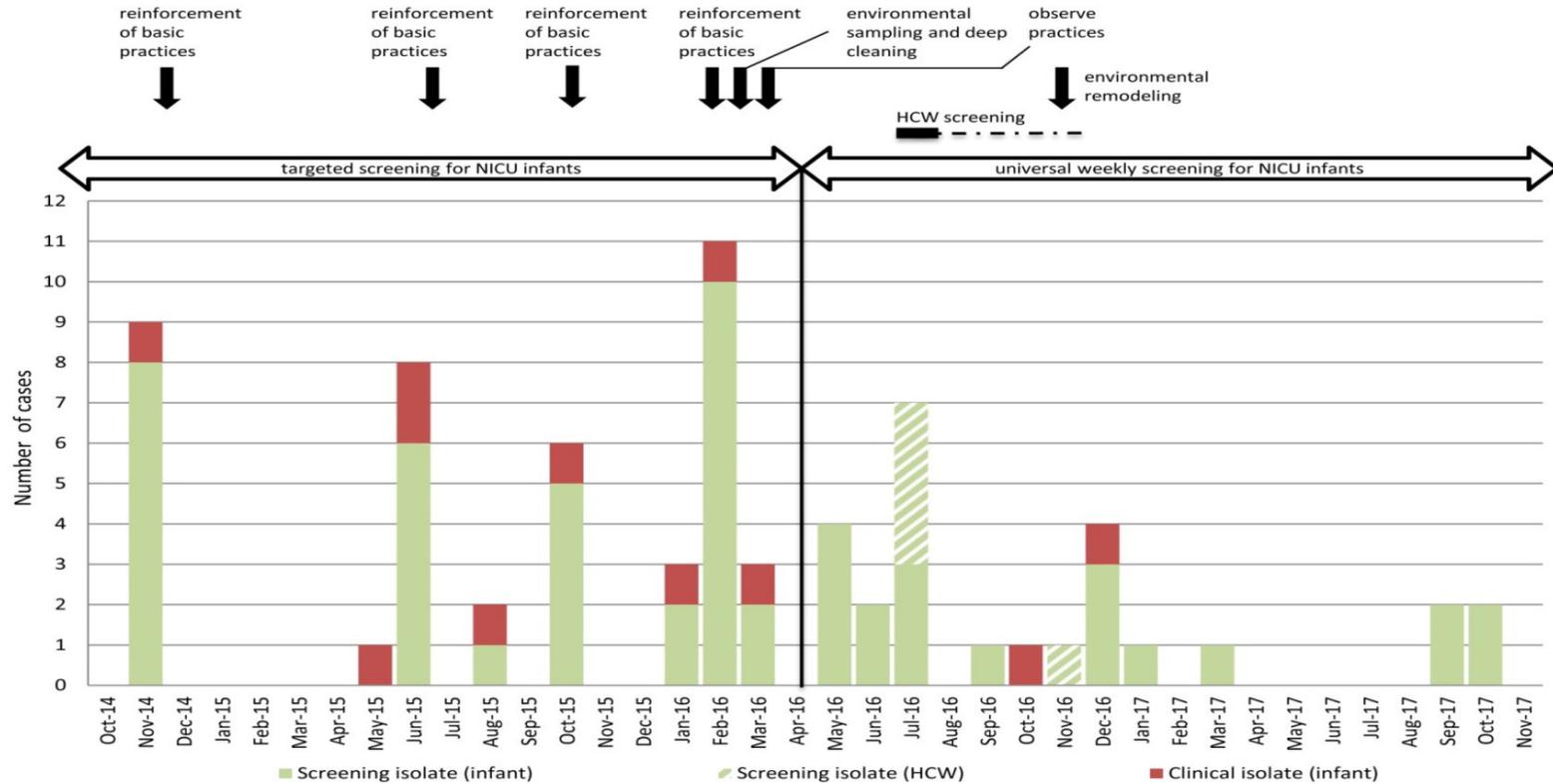


1982-2020

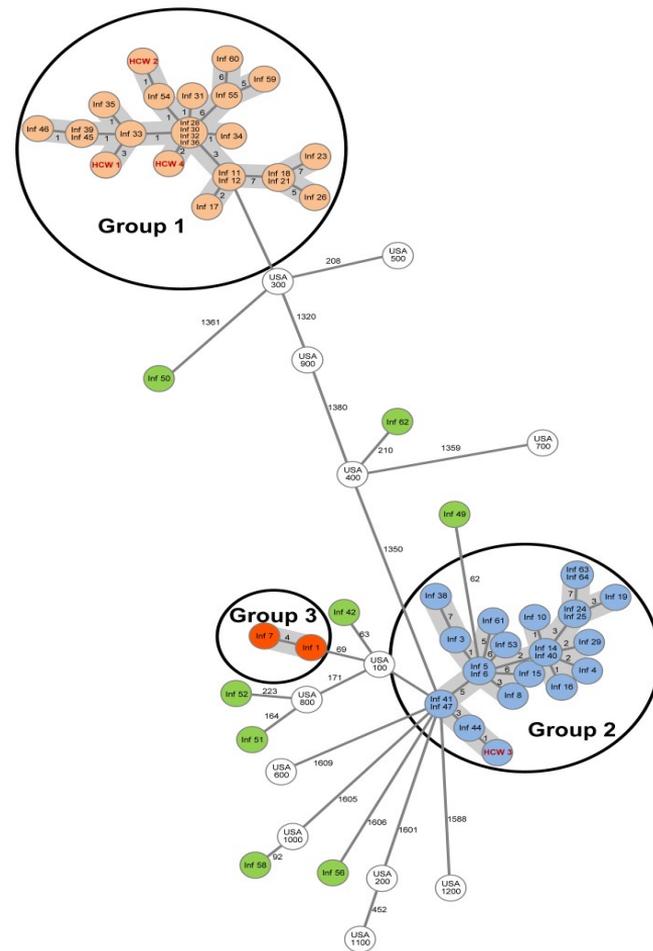


Technology Revolution: Whole Genome Sequencing

Neonatal Intensive Care Unit MRSA Cases by Initial Source & Interventions Taken

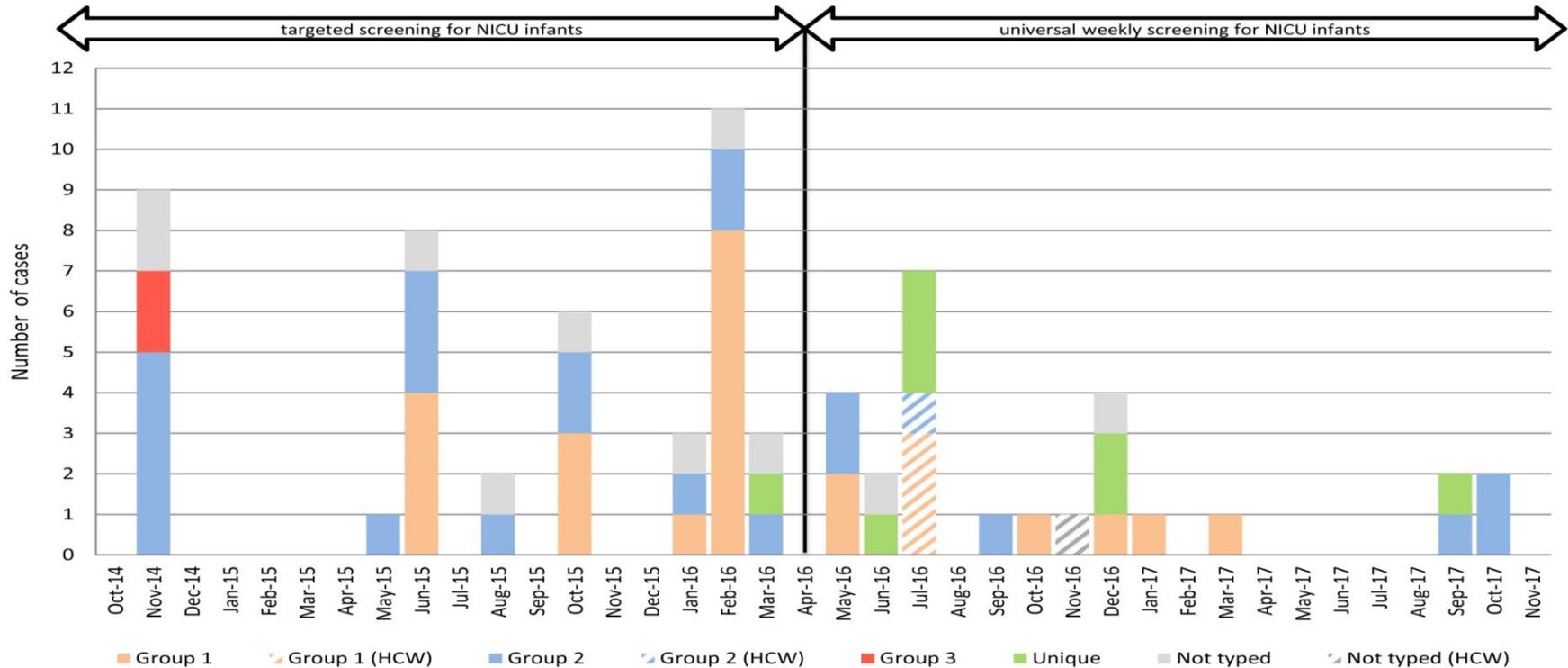


Neonatal Intensive Care Unit Isolates (Clinical & Surveillance) Neonates & Healthcare Workers



NICU Outbreaks (!) Timeline

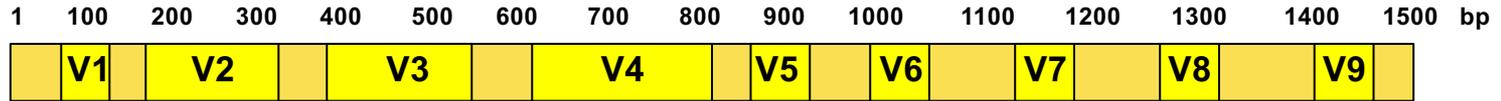
MRSA Cases Shown by WGS Group



Species Validated for WGS (cgMLST)

- *Acinetobacter baumannii*
- *Campylobacter jejuni/coli*
- *Clostridioides difficile*
- *Cutibacterium acnes*
- *Enterobacter cloacae*
- *Enterococcus faecalis*
- *Enterococcus faecium*
- *Escherichia coli*
- *Klebsiella pneumoniae*
- *Legionella pneumophila*
- *Pseudomonas aeruginosa*
- *Serratia marcescens*
- *Staphylococcus aureus*
- *Staphylococcus epidermidis*
- *Staphylococcus lugdunensis*
- *Streptococcus agalactiae*
- *Streptococcus pyogenes*

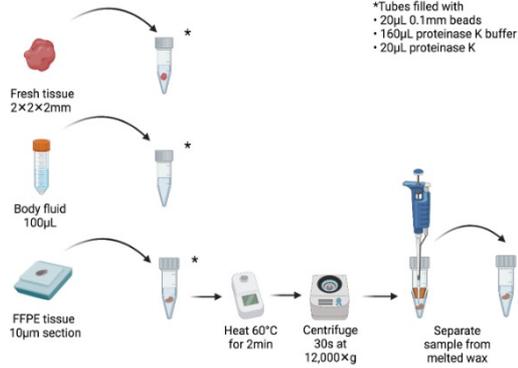
Technology Revolution: Targeted Metagenomic Sequencing (16S Ribosomal RNA Gene)



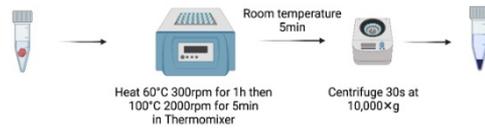
16S rRNA Gene PCR/Sanger Sequencing in Clinical Practice

- Retrospective analysis specimens clinically analyzed by 16S rRNA PCR/Sanger sequencing (04-2017→ 03-2019)
- 566 specimens, 460 patients
- 17% (97/566) 16S rRNA gene PCR/Sanger sequencing positive
 - 90% (88/97) positive results → clinical infections
- 22% positive specimens (21/97), 16S rRNA gene detected but bacterium not identified by Sanger sequencing (mixtures)

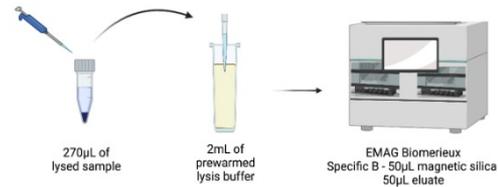
1 Sample processing



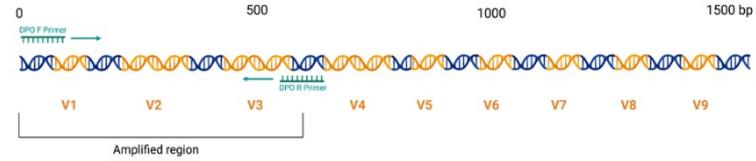
2 Digestion and lysis



3 Extraction



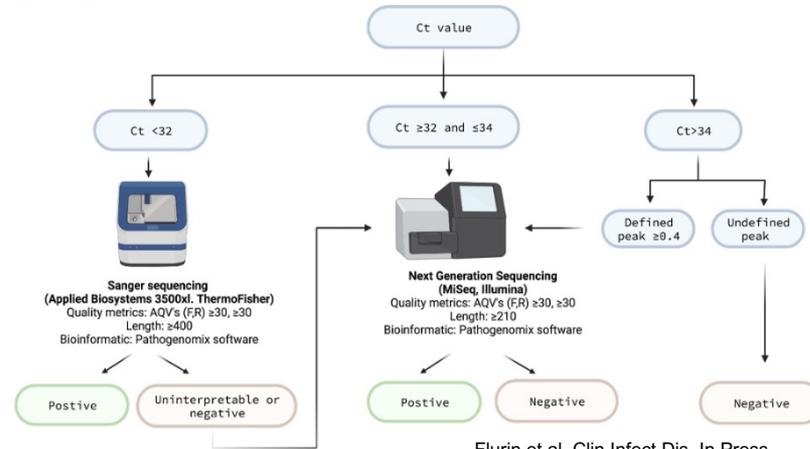
4a 16S rRNA gene and primers



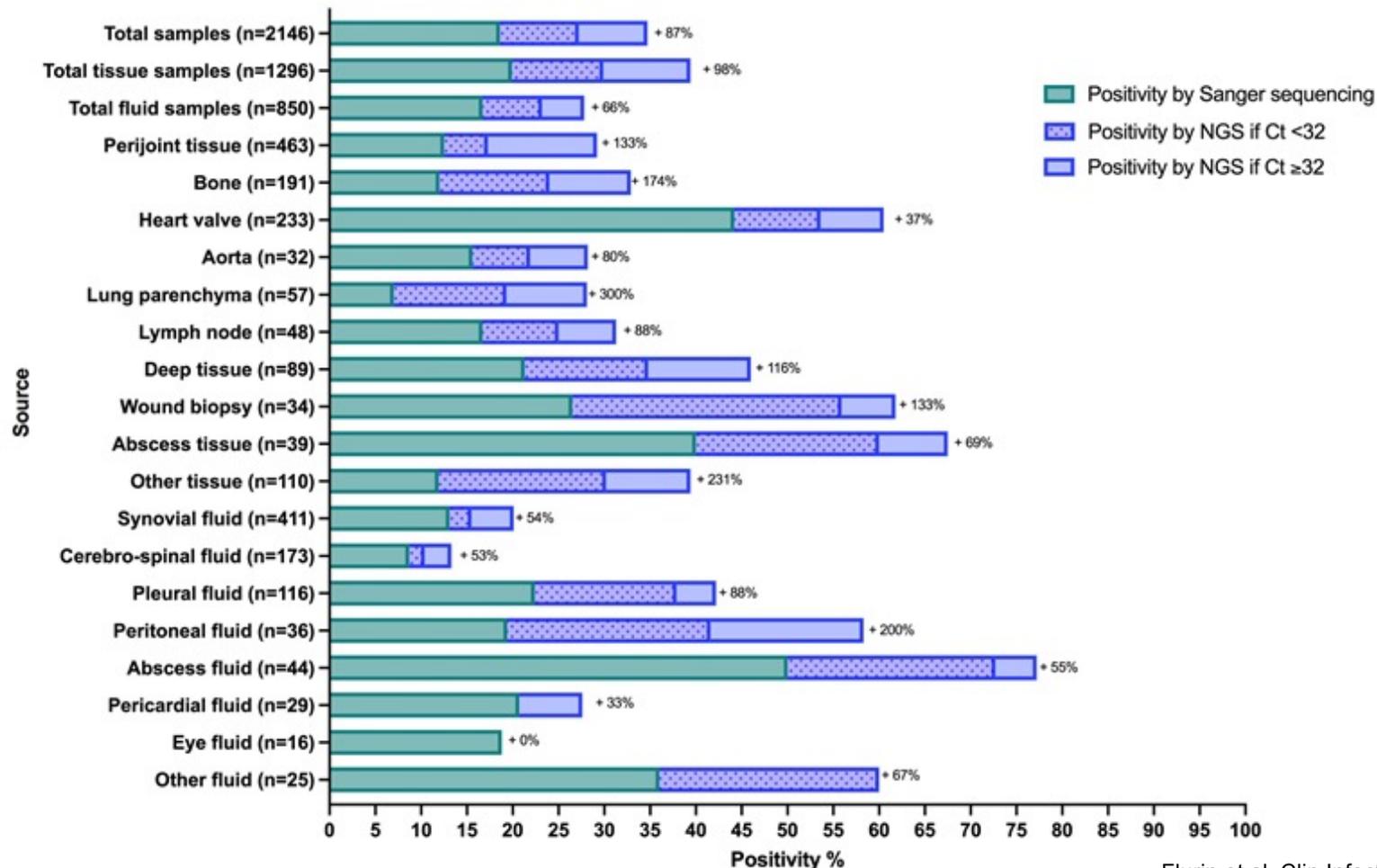
4b 16S rRNA PCR



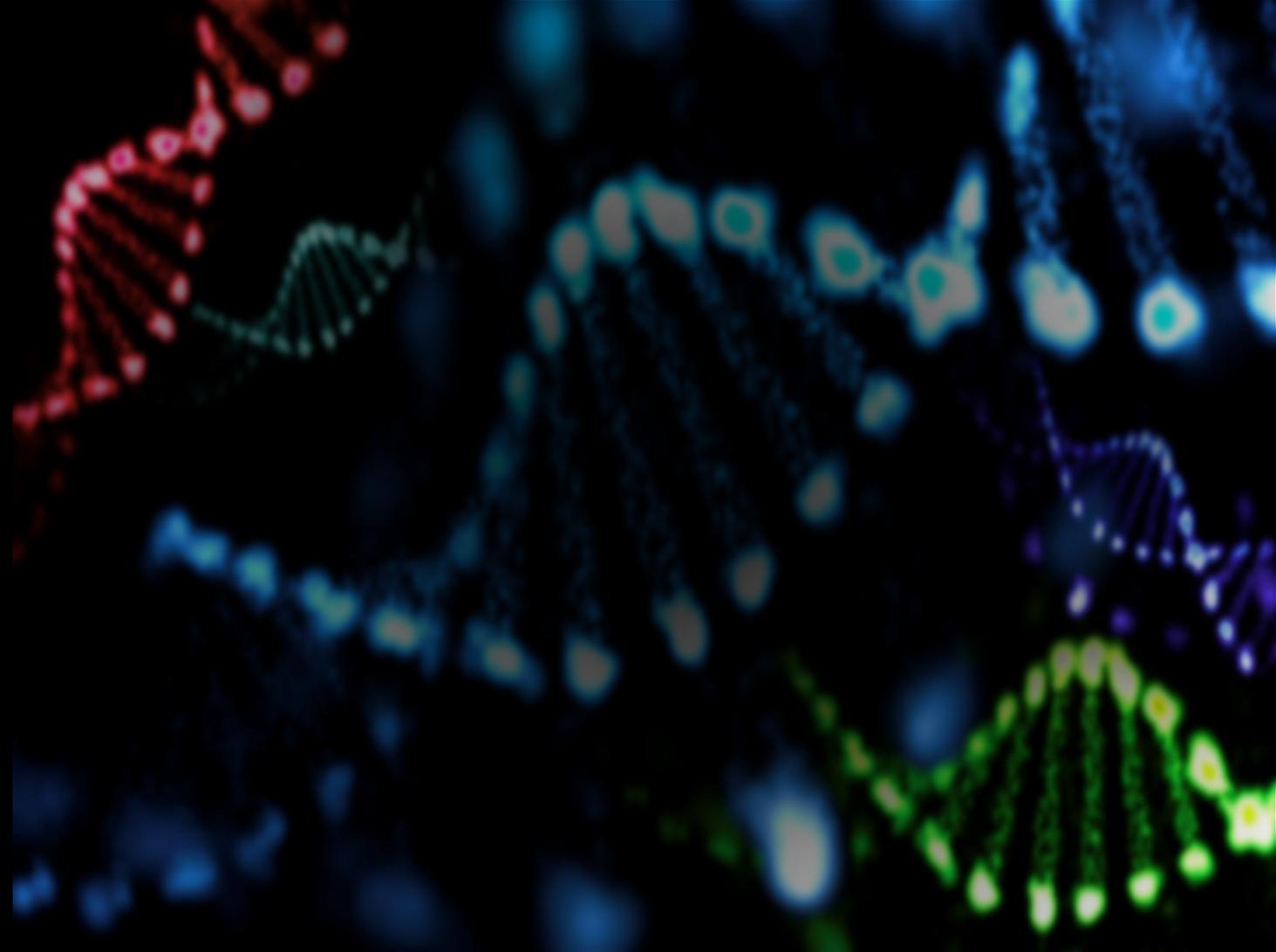
5 Sequencing algorithm



Yield of next-generation sequencing (NGS)



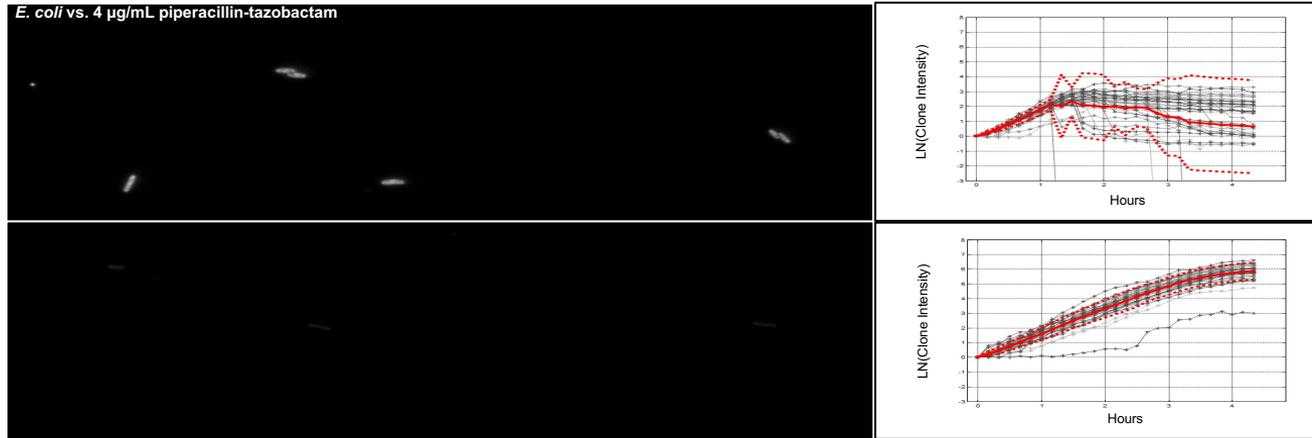




Technology Revolution: Single Cell Imaging

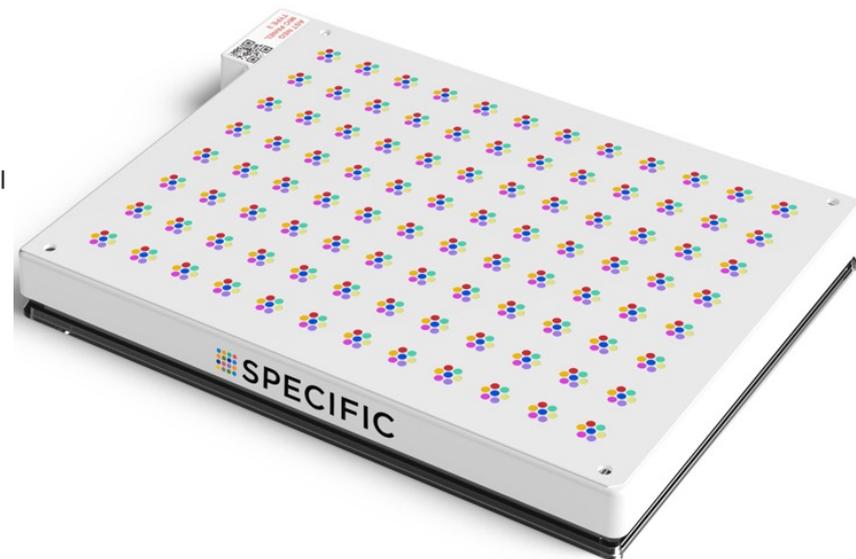
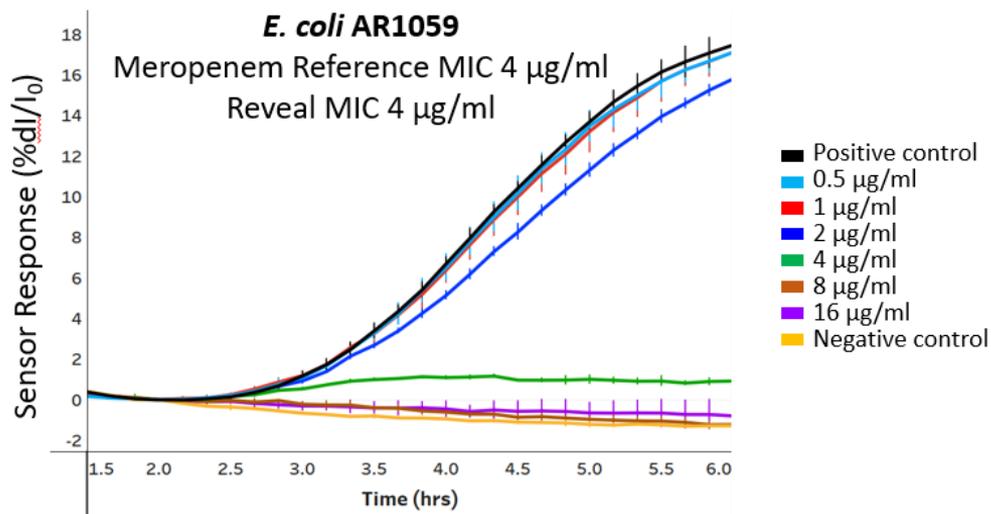
Application: Antibacterial Susceptibility Testing

- Time-lapse darkfield imaging, analysis of bacterial growth
 - Individual bacterial response to single antibiotic concentration over time



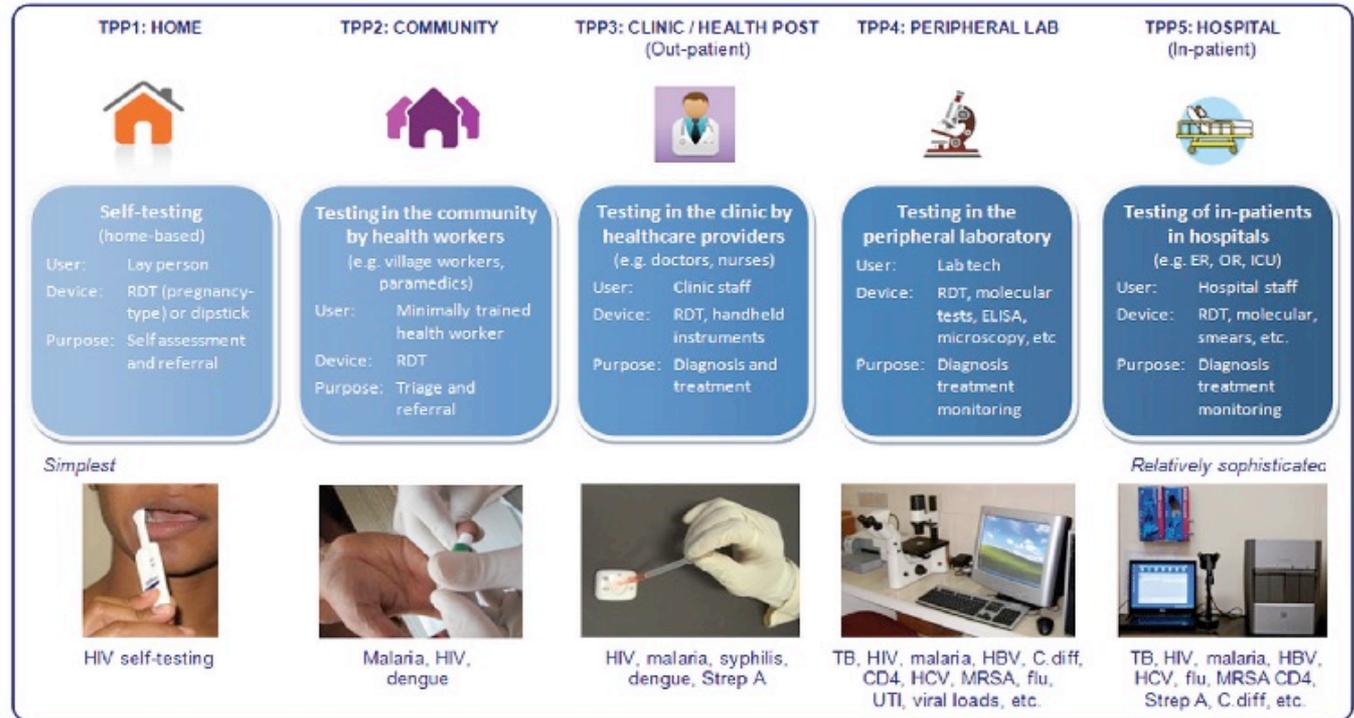
Technology Revolution: Metabolomics

Application: Antibacterial Susceptibility Testing



Technology Revolution

Point of Care Diagnostics



www.asm.org/index.php/colloquium-reports/item/6421-changing-diagnostic-paradigms-for-microbiology

Technology Revolution: Advanced Host Response Assessment for Infectious Diseases

- **Advanced human transcriptomic and/or proteomic analyses may be able to**
 - Point to underlying viral, bacterial, fungal, parasitic infections and non-infectious processes
 - Possibly, point to specific microbial etiologies
 - Blood, body fluids, tissues?

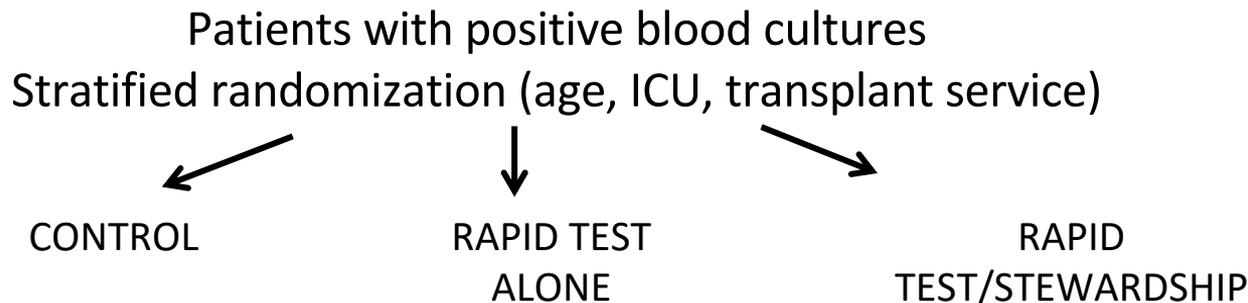
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How to Move to the Future?

- **MALDI-TOF mass spectrometry example**
- **Demonstration of clinical utility**
 - COVID-19 example
 - Advanced diagnostics - sometimes not so clear...

Randomized Controlled Clinical Trial BioFire BCID, Mayo Clinic 2013-2014



- Gram stain called to service
- Standard subculture and susceptibility (1-3 d)

- Gram stain called to service
- Standard subculture and susceptibility (1-3 d)
- Rapid test plus lab call with comments (1 h)

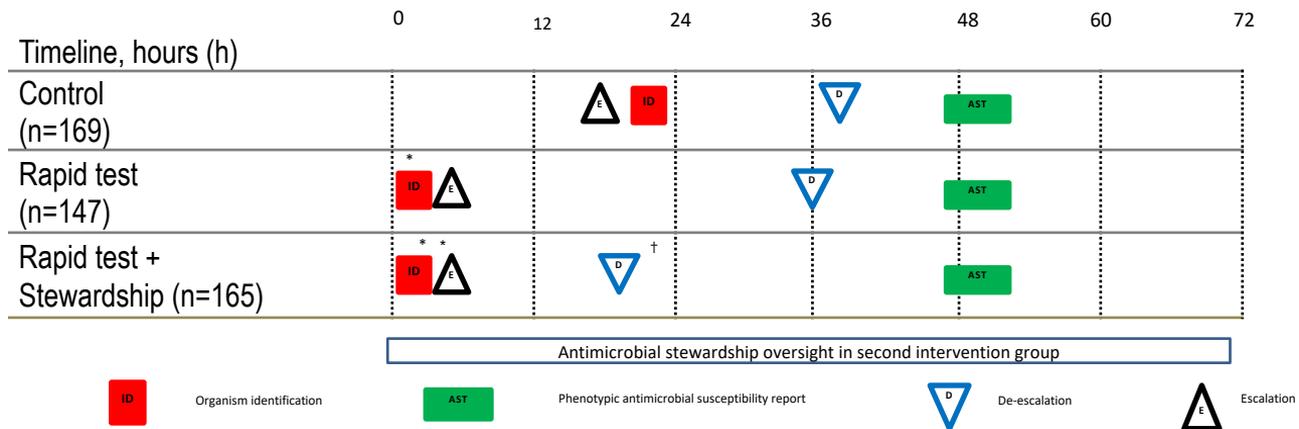
- Gram stain called to service
- Standard subculture and susceptibility (1-3 d)
- Rapid test plus lab call with comments (1 h)
- ID MD/pharmacist call with specific treatment recommendations

Banerjee et al. Clin Infect Dis 2015;61:1071

Supported by the National Institute of Allergy And Infectious Diseases of the National Institutes of Health under Award Number UM1AI104681 (Antibacterial Resistance Leadership Group)



Comparison of Time To Identification, Susceptibility Results, and Antibiotic Modifications



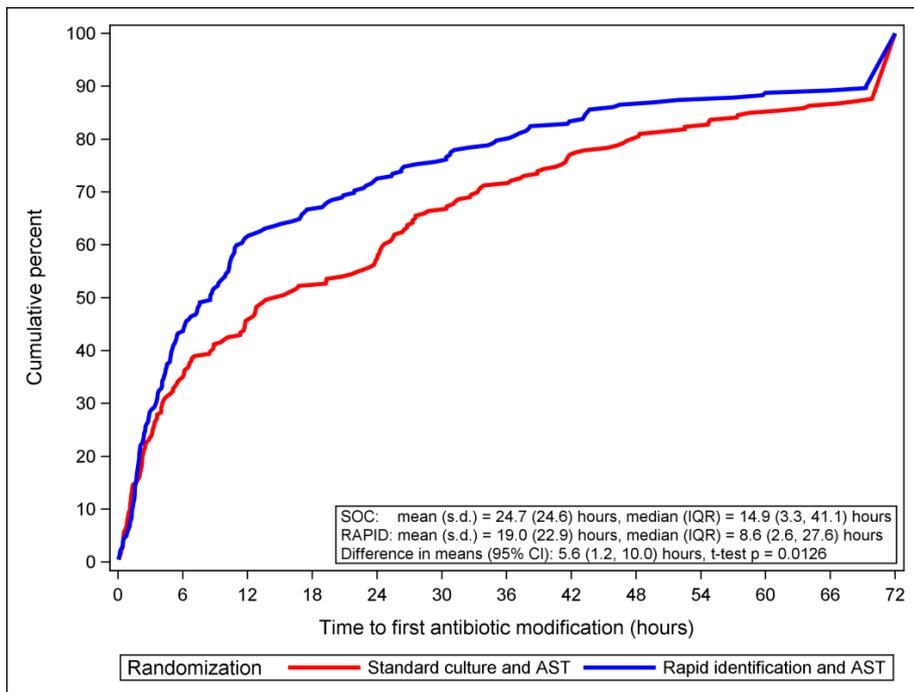
*Significant vs. control; †Significant vs. control and rapid multiplex PCR alone

Thoughtful, Clear and Informative Test Reporting

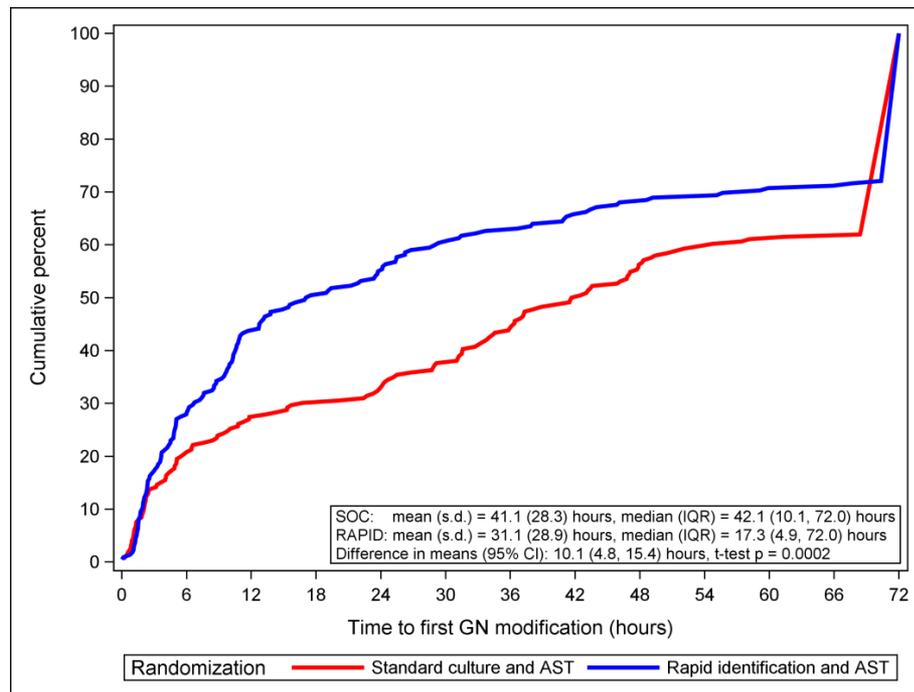
| FilmArray BCID2 Result | Report | Isolate Comments |
|---|--------------------------------------|--|
| <i>Staphylococcus aureus</i> + <i>mecA/C</i> and MREJ (MRSA) + | <i>Staphylococcus aureus</i> complex | <i>mecA</i> detected Methicillin (oxacillin)-resistant <i>Staphylococcus aureus</i> complex (MRSA). MRSA is predictably resistant to beta-lactam antibiotics (except ceftaroline). Vancomycin or other anti-MRSA treatment recommended for initial therapy pending susceptibility results. Semi-Urgent Result Consult Infectious Diseases |
| <i>Staphylococcus aureus</i> + <i>mecA/C</i> and MREJ (MRSA) - | <i>Staphylococcus aureus</i> complex | <i>mecA</i> not detected Methicillin (oxacillin)-susceptible <i>Staphylococcus aureus</i> complex. An anti-staphylococcal beta-lactam, such as cefazolin, nafcillin or oxacillin, is recommended for empiric therapy, unless clinically contraindicated. Consult Infectious Diseases |
| <i>Staphylococcus epidermidis</i> + <i>mecA/C</i> + | <i>Staphylococcus epidermidis</i> | <i>mecA</i> detected Methicillin (oxacillin)-resistant <i>Staphylococcus epidermidis</i> . Possible blood culture contaminant (unless isolated from more than one blood culture draw or clinical case suggests pathogenicity). No antibiotic treatment is indicated for blood culture contaminants. |

Randomized Trial Evaluating Clinical Impact of RAPid Identification and Susceptibility Testing for Gram-negative Bacteremia: RAPIDS-GN

Time to any antibiotic change

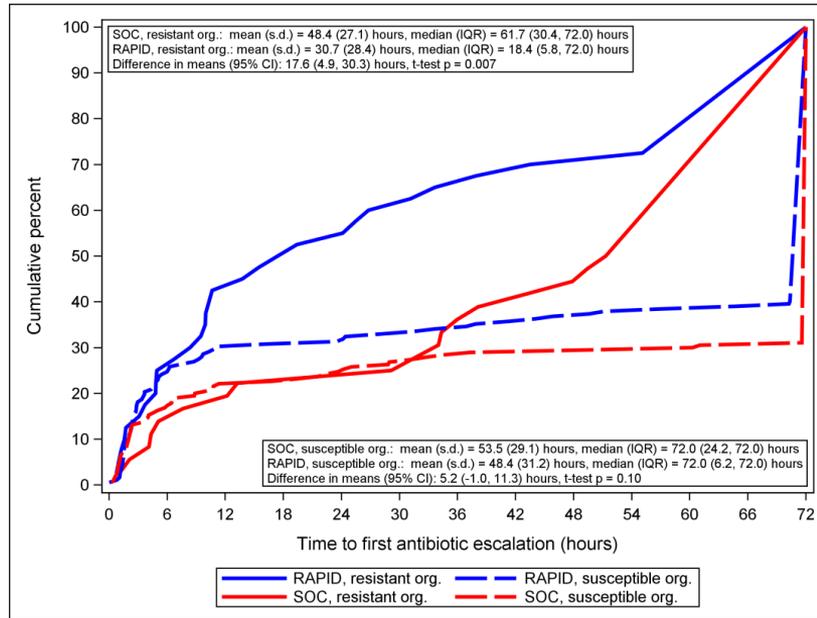


Time to GN antibiotic change

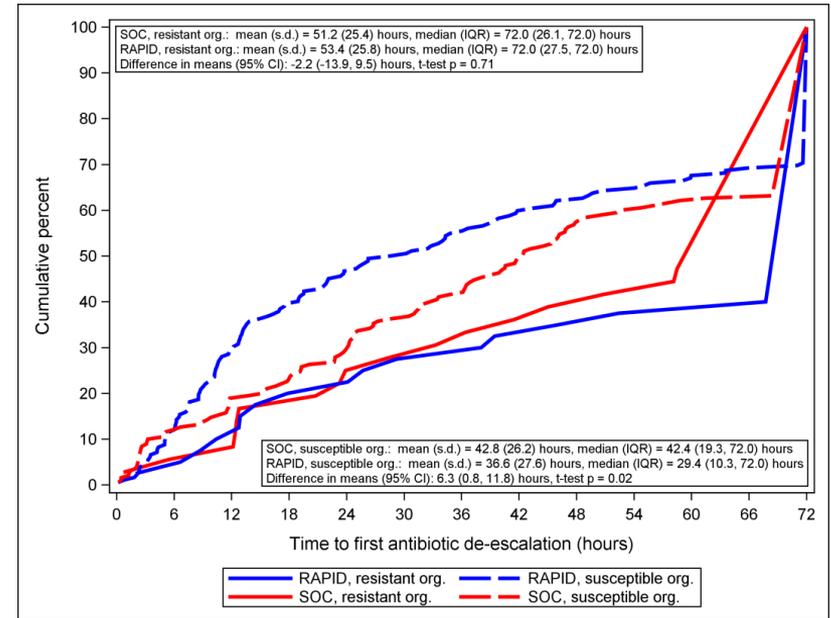


Randomized Trial Evaluating Clinical Impact of RAPID Identification and Susceptibility Testing for Gram-negative Bacteremia: RAPIDS-GN

Time to antibiotic escalation

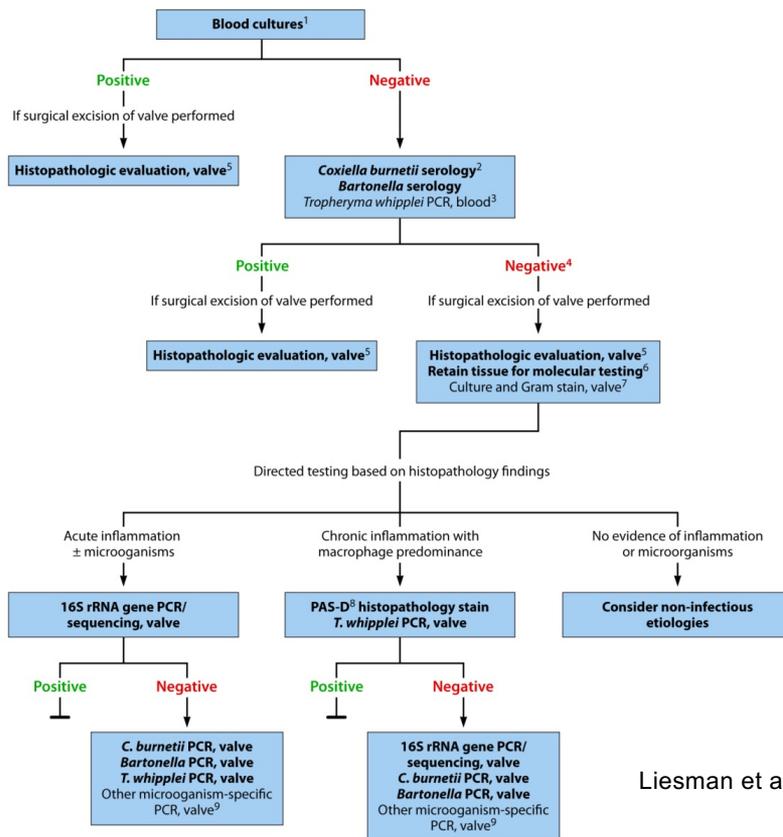


Time to antibiotic de-escalation



Importance of Guidelines, Diagnostic Algorithms, Diagnostic Stewardship

Microbiologic and Pathological Diagnostic Algorithm For Endocarditis



Liesman et al J Clin Microbiol. 201;55(9):2599-608

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The Deliverables: Improved Patient Health

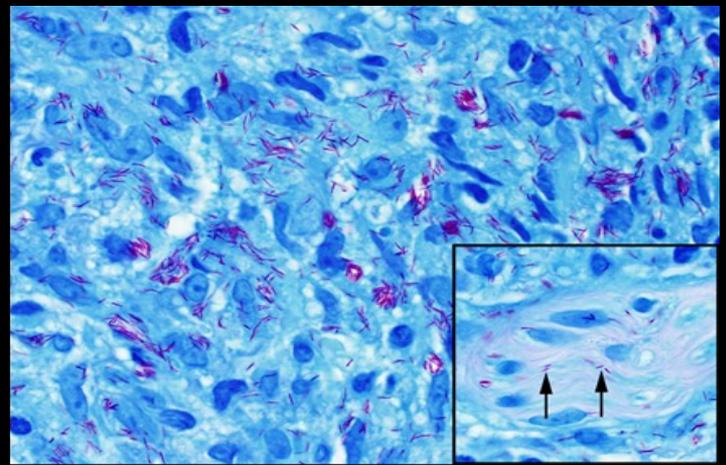
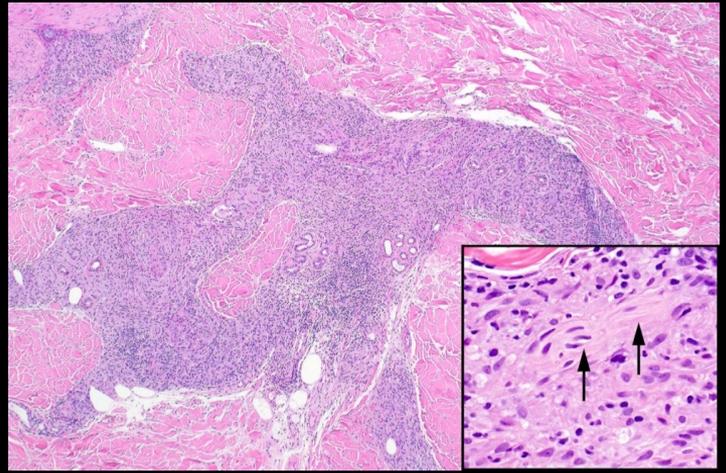
- **Improved patient health and hopefully...**
 - Avoid unneeded testing and treatment (patient benefit)
 - Decrease transmission of infectious diseases (societal benefit)
 - Curb emergence of antimicrobial resistance (patient and societal benefit)

The Deliverables: Changes in Healthcare Delivery

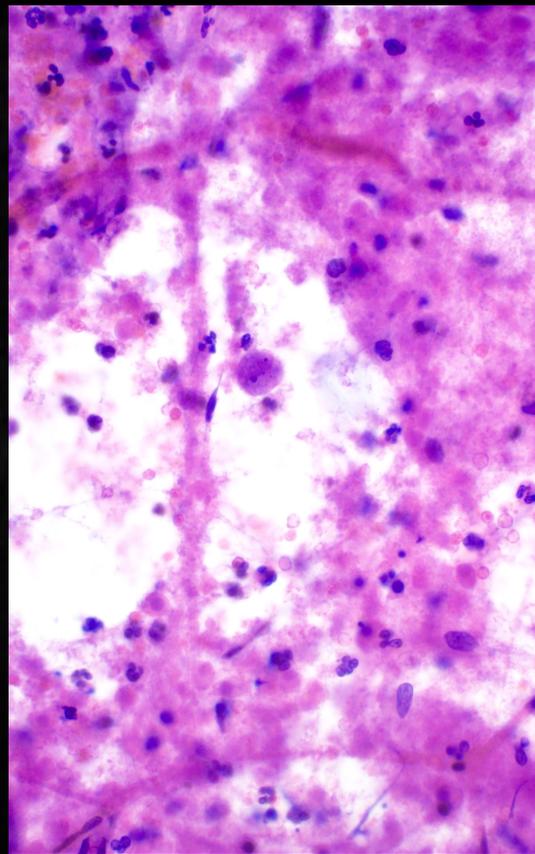
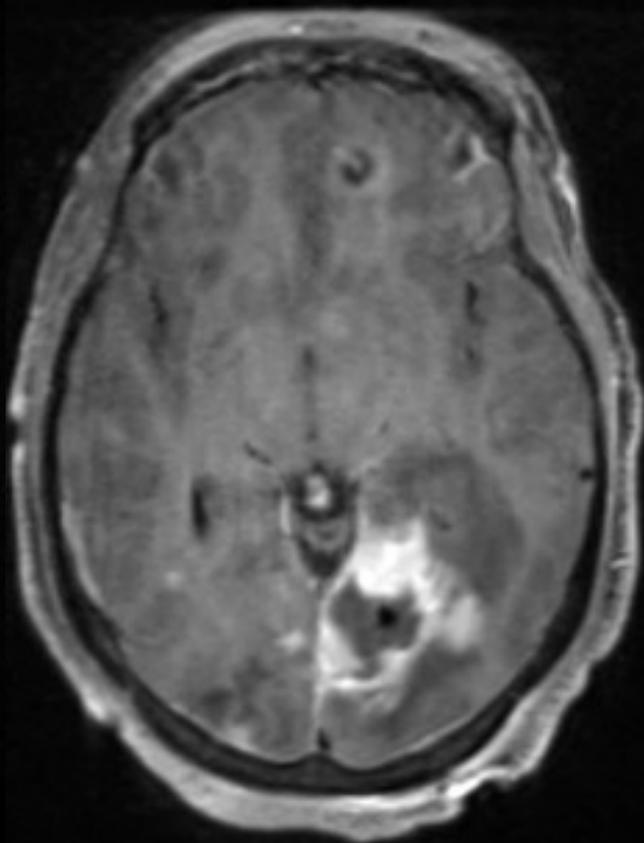
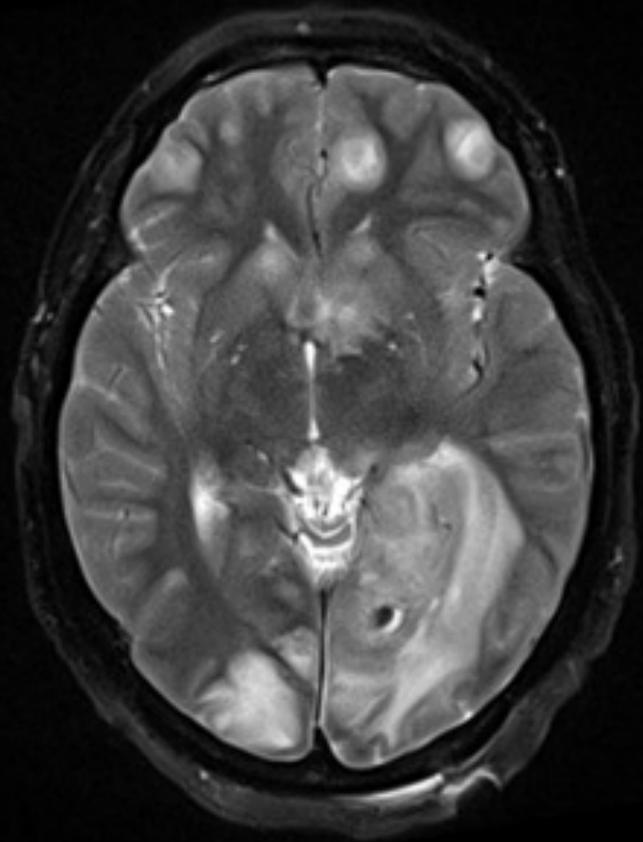
- **Home-based testing**
- **Patient self-testing**
- **Testing @ non-traditional sites**
- **Test prioritization approaches in traditional healthcare sites (e.g., emergency departments, urgent care settings)**
- **etc.**

- **Discovery through testing**

- *Borrelia mayonii*, *E. muris* subspecies *eauclairensis*, *Yersinia rochesterensis*
 - Many more novel species remain to be described
- Understanding antimicrobial resistance
 - Characterization of vancomycin resistance in *Paenibacillus popilliae*
 - Recognition of the frequency of resistance in *Helicobacter pylori*
- New diseases – hyperammonemia associated with *Ureaplasma* species



| | | | |
|----------------|--|---|------|
| Source: | BRAIN, RIGHT- Tissue. Rlght frontal lobe | | |
| Fresh vs FFPE: | Fresh | | |
| Cp value: | 31.86 | | |
| FORWARD | | | |
| Abundance | Read length | Result | % ID |
| | 22606 | 222Amoebophilus | 96.8 |
| | 405 | 225Cutibacterium acnes ** | 100 |
| | 274 | 224Methylothera | 95.5 |
| | 222 | 222Amoebophilus | 96.4 |
| | | Chryseobacterium lactis / Chryseobacterium aquaticum / Chryseobacterium aurantiacum / | |
| | 211 | 223Chryseobacterium vietnamense | 99.1 |
| | 148 | 224Amoebophilus | 96.4 |
| | 140 | 222Enterococcus faecium / Enterococcus hirae / Enterococcus pseudoavium | 100 |
| | 138 | 222Amoebophilus | 96.8 |
| | 128 | 222Amoebophilus | 96 |
| | | | |
| | 117 | 222Amoebophilus | 96.4 |
| | 106 | 222Amoebophilus | 96.4 |
| REVERSE | | | |
| | 19971 | 228Amoebophilus | 98.2 |
| | 754 | 229Amoebophilus | 98.3 |
| | 353 | 226Cutibacterium acnes / Cutibacterium namnetense * | 100 |
| | 179 | 227Methylophilus | 98.7 |
| | 170 | 228Amoebophilus | 97.8 |
| | 144 | 228Amoebophilus | 97.8 |
| | | Enterococcus durans / Enterococcus faecium / Enterococcus hirae / Enterococcus ratti / | |
| | | Enterococcus thailandicus / Enterococcus villorum / Enterococcus xinjiangensis / Enterococcus | |
| | 139 | 227lactis / Enterococcus pseudoavium | 100 |
| | | Chryseobacterium artocarpi / Chryseobacterium antibioticum / Chryseobacterium carnipullorum | |
| | 131 | 228/ Chryseobacterium shigense / Chryseobacterium ureilyticum / Chryseobacterium vrystaatense | 99.6 |
| | 118 | 228Amoebophilus | 97.4 |
| | 118 | 228Amoebophilus | 97.4 |
| | 104 | 228Amoebophilus | 97.8 |



Conclusions

- **Many current microbiology diagnostics antiquated, poorly used**
- **Technology revolution**
 - Bringing major advances
- **How to move to the future**
 - Demonstration of value
- **The deliverables**
 - Improved health (patients, society, antimicrobial resistance)
 - Changes to healthcare delivery
 - Better understanding of infectious diseases and their mimics (discovery through testing)



R01 AR056647
R01 AI91594
R21 AI125870
UM1 AI104681